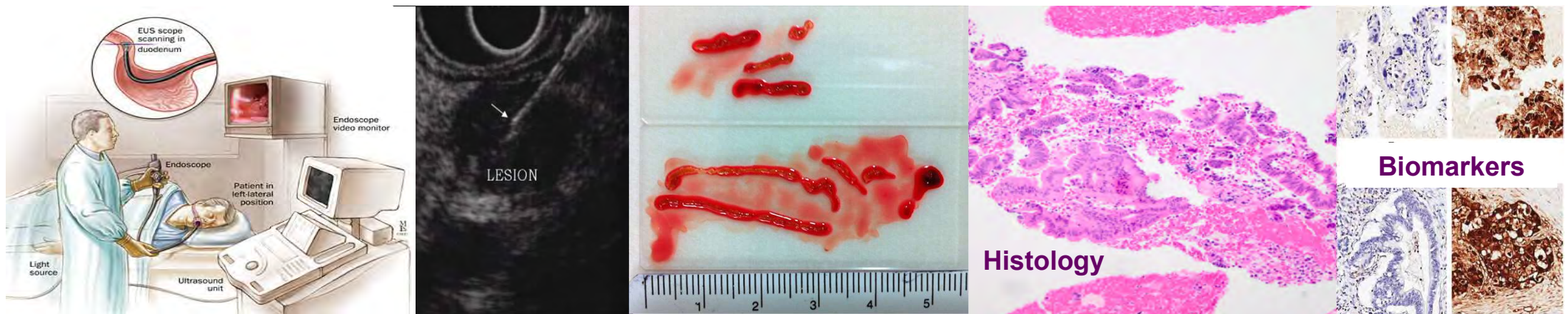


# EUS guided FNA of GI diseases

## Challenges for endoscopists and pathologists

*Do we really need rapid on-site assessment?*



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Member of the Australian Pancreatic Cancer Genome Initiative

Member of the International Cancer Genome Consortium

Discipline of Medicine, University of Adelaide, South Australia

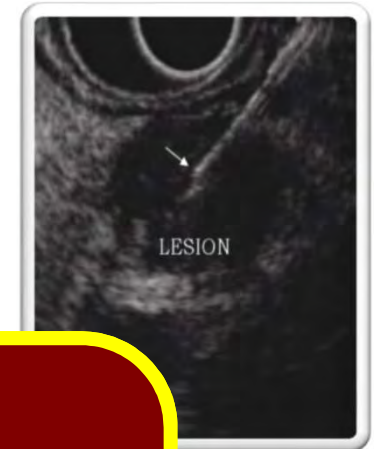
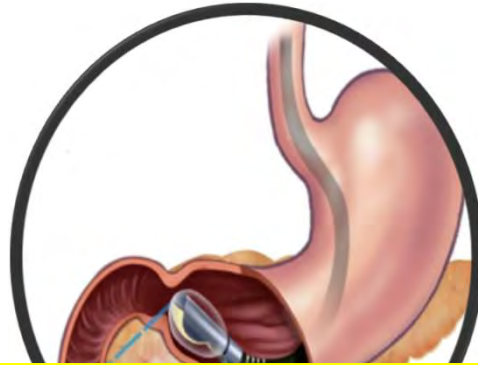
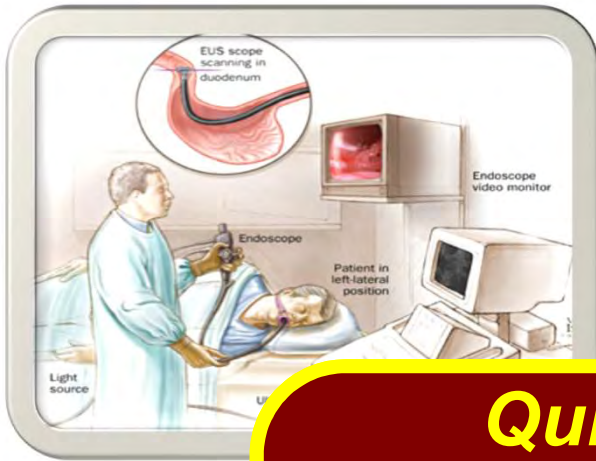


# Outline

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- *Challenges in EUS FNA for endoscopists*
- *Challenges in EUS FNA for pathologists*
- *Impact of Rapid Onsite Cytological Evaluation (ROSE) for the endoscopists and pathologist*
- *Alternative approach if ROSE is not presence*
  - Does cytology still needed?
  - Macroscopic Onsite (core) Evaluation (MOSE)
  - Routine core needle (FNB) biopsy

# ***Current status of EUS guided FNA***



***Quick, easy to use, and safe***

***Diagnostic outcome:***

- Sensitivity = 76-98%***
- Specificity = 60-100%***
- PPV = 93-100%***



# EUS-guided FNA is better than percutaneous approach for GI and Pancreatico-biliary neoplasm



Diagnostic yield from needle biopsy is higher with EUS guided approach:

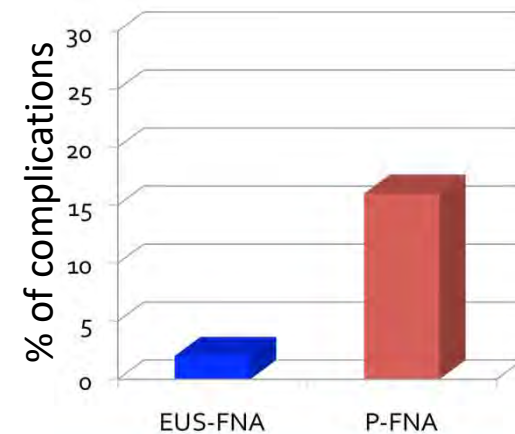
- **EUS-FNA:** 84%
- CT / US FNA: 62%

*Horwhat et al. GIE 2006*

Peritoneal carcinomatosis at time of surgery

|                          |            |
|--------------------------|------------|
| Percutaneous FNA (P-FNA) | 16 %       |
| <b>EUS-FNA</b>           | <b>2 %</b> |

*Micames et al. GIE 2003*



# EUS guided tissue acquisition is ....an ART!

**No single “gold standard” needle, technique or scope position!**

**Best outcome and the choice of needle, technique and tissue processing will depends on may factors:**

- Location and size of lesion
- Indication of procedure
  - Need for biomarkers or genomics analysis
- Presence of ROSE
- Inability to stop anti-platelet or anti-coagulant therapy
- Endoscopist's experience (*this will overcome all others!*)

# Needle and technique factors in EUS FNA outcomes

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- **Type of needle**

- Size
- Flexibility
- Shape of needle tip

## **Techniques**

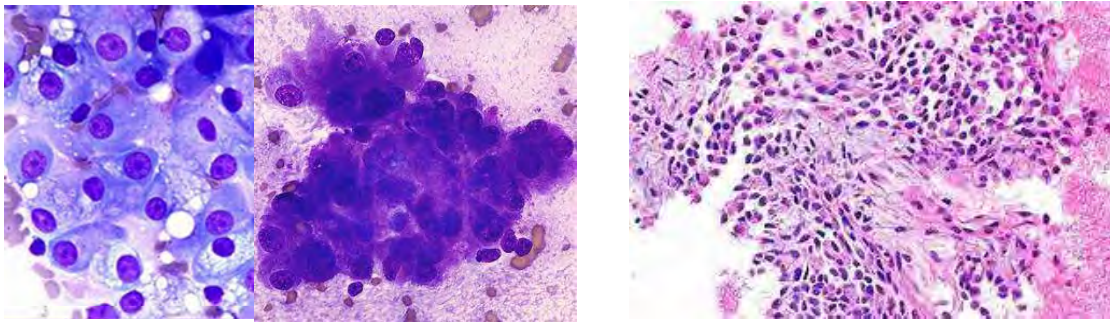
- Number of pass
- Stylet
- Suction, slow pull vs. none
- “Fanning”
- Ancillary imaging techniques to guide the site of biopsy
  - Elastography
  - Contrast
- Site of needle puncture

# Type of specimens from different needles

## Sharp tip needles

Small calibre (25G/22G)  
Highly flexible and easy to use  
Tiny to small amount of tissue  
Suitable for cytological evaluation

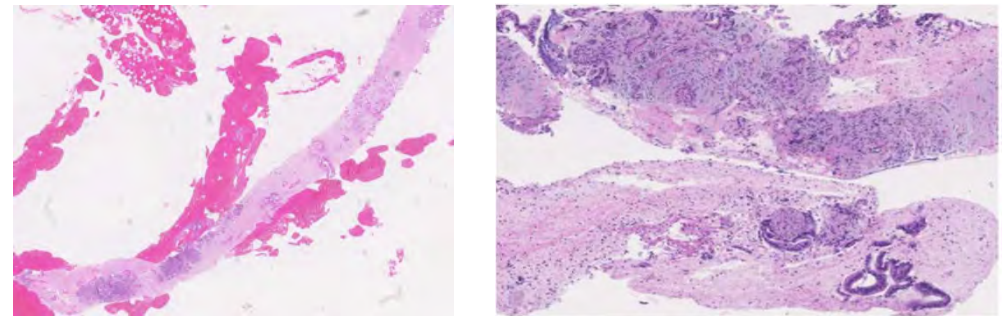
## FNA needles



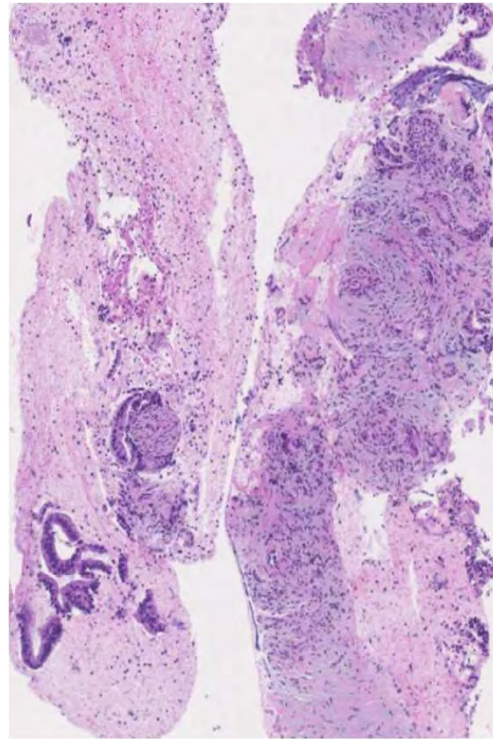
## Modified tip needles

- Medium calibre (22G/20G/19G)
- Less flexible and sharp
- Harder to use
- Acquire core tissue
- Suitable for histology

## FNB needles



# EUS Tru-cut needle



**Good core tissue**

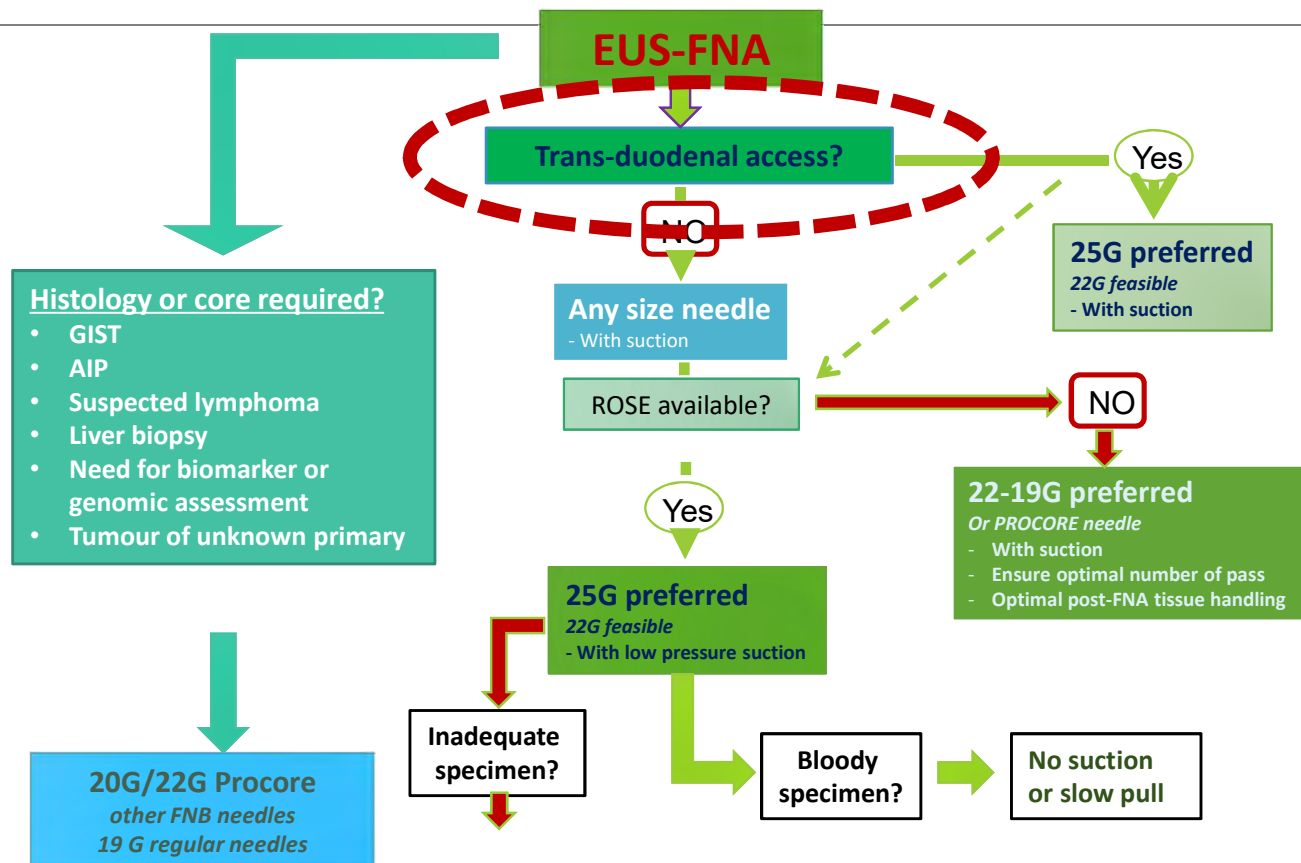
**Compromised by:**

- Technically difficult with high failure rate
- Extremely rigid
- Almost impossible for lesions located in pancreatic head or uncinate process

**No longer use, and is replaced by the newer tip-modified needle.**



# Algorithm that based on scope position to determine type and size of needle...



# Number of pass?

*Relevant only when ROSE is not available*

**Recommend performing 3 needle passes for lymph nodes and liver lesions, at least 5 needle passes for solid pancreatic masses (mostly used 22G)**

- *Pancreatic lesion: Four passes of 25G is sufficient?.*

*Suzuki et al. Dig Endosc 2012*

## **Less for Core (FNB) needle**

- **Single-pass studies: 87% for 19G , 90% for 20G, 88% for 22G Procore**

*Iglesias-Garcia et al. Endoscopy 2011   Larghi et al. Endoscopy 2014   Larghi et al. EuroEUS 2015*

- **Even with ROSE, 22G Procore required fewer passes than FNA**

- *1 vs. 2 passes; P<0.0001;*
- *Procore had higher % diagnosis on 1<sup>st</sup> pass: 73% vs. 37%; P<0.001*

*Lee et al. Endoscopy 2014*

# Technique: Fanning?



N=54, ROSE, Pancreatic mass  
Randomised:

- “Fanning” = 26 vs. Standard = 28

Results:

## ***Diagnostic accuracy***

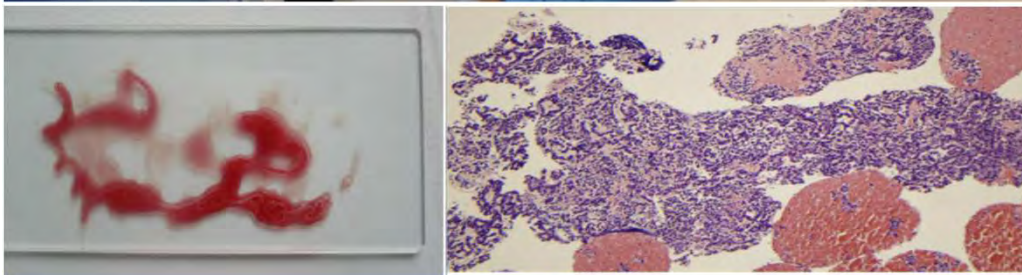
- 96% vs. 77% (P=0.05)

***Lesser no. pass*** required for diagnosis  
and ***higher % of diagnosis on 1<sup>st</sup> pass***

- 86% vs. 58% (P=0.02)

***Bang et al. Endoscopy 2013; 45; 445***

# Slow pull or capillary suction



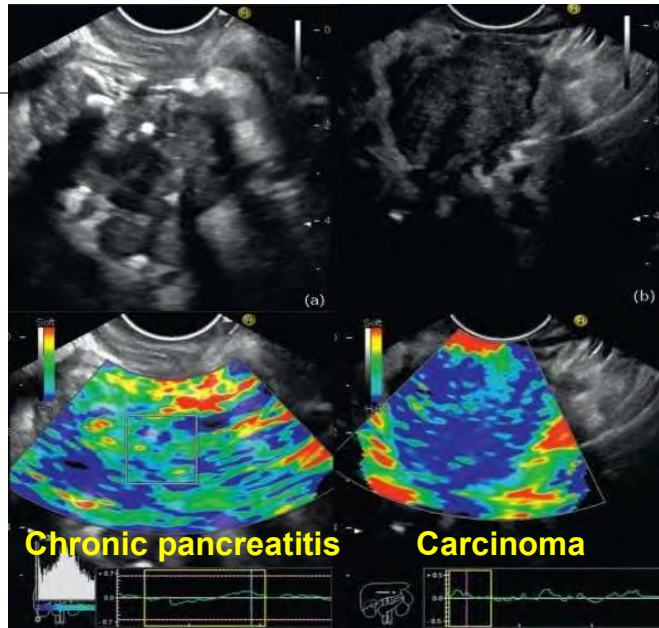
## *Nakai et al. Dig Dis Sci 2014*

- Retrospective study, pancreatic mass
- 181 suction vs. 186 slow pull FNA
- Both 25G and 22G were examined.

### **Results:** Slow pull is associated with

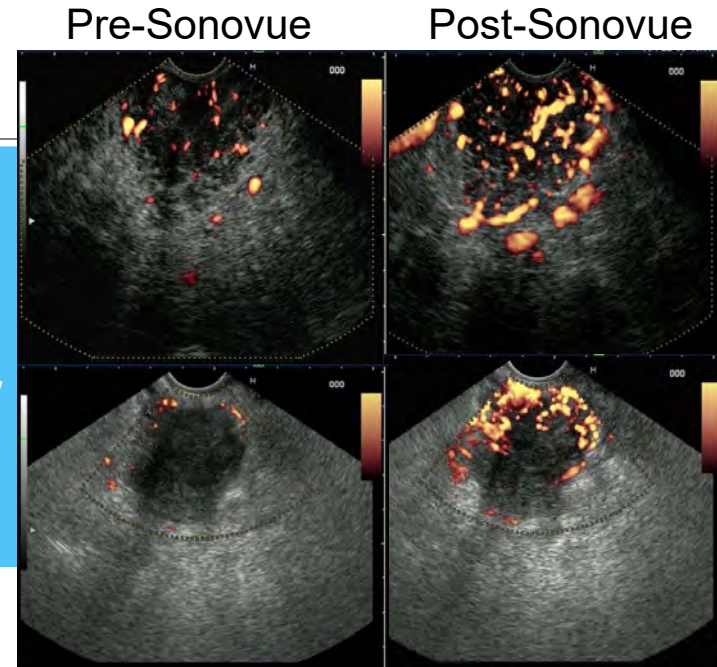
- Less blood contamination but also less cellularity but higher diagnostic yield only with 25G?
- No impact with 22G

# Advanced imaging assisted in difficult cases

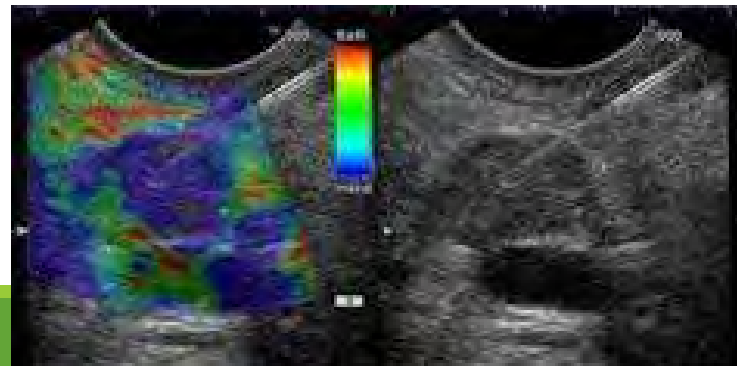


*Iglesias-Garcia et al. Gastroenterology 2010*

Predict the likelihood of malignancy prior to FNA



Guide placement of needle during FNA



***“Only blood...”***

***“No diagnostic tissue...”***

***“Atypical cells but non-diagnostic”***

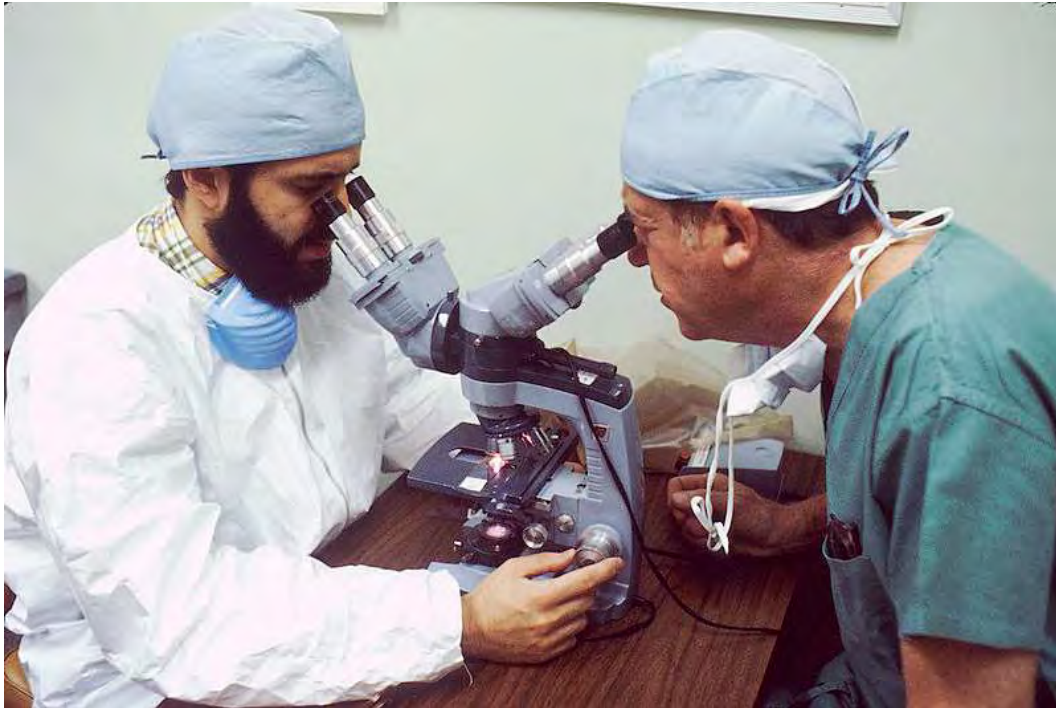
***“Insufficient material for diagnosis”***

*“Atypical cells, highly suspicious of malignant process”*

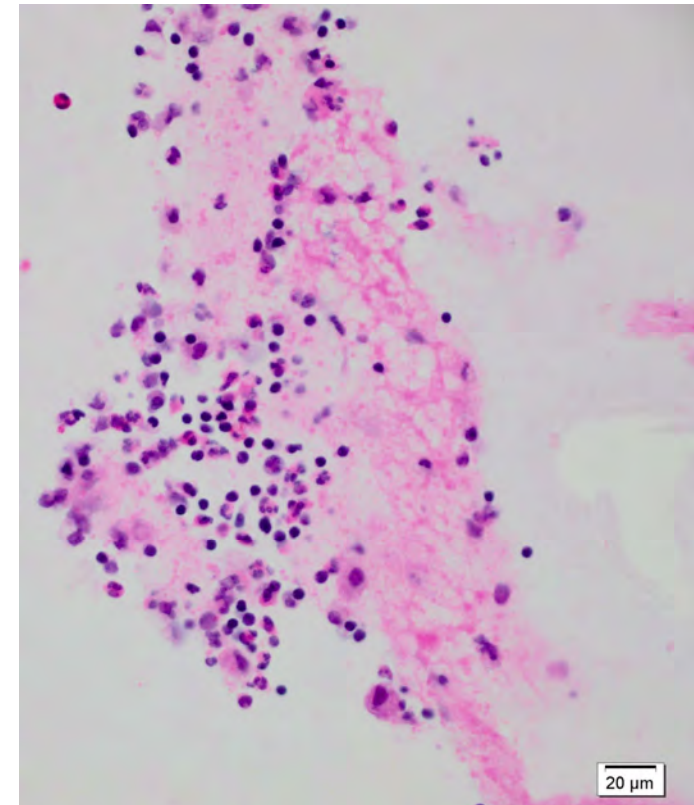


# How to overcome the endoscopist's frustration?

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# EUS FNA and Rapid On-site Cytology Evaluation (ROSE)





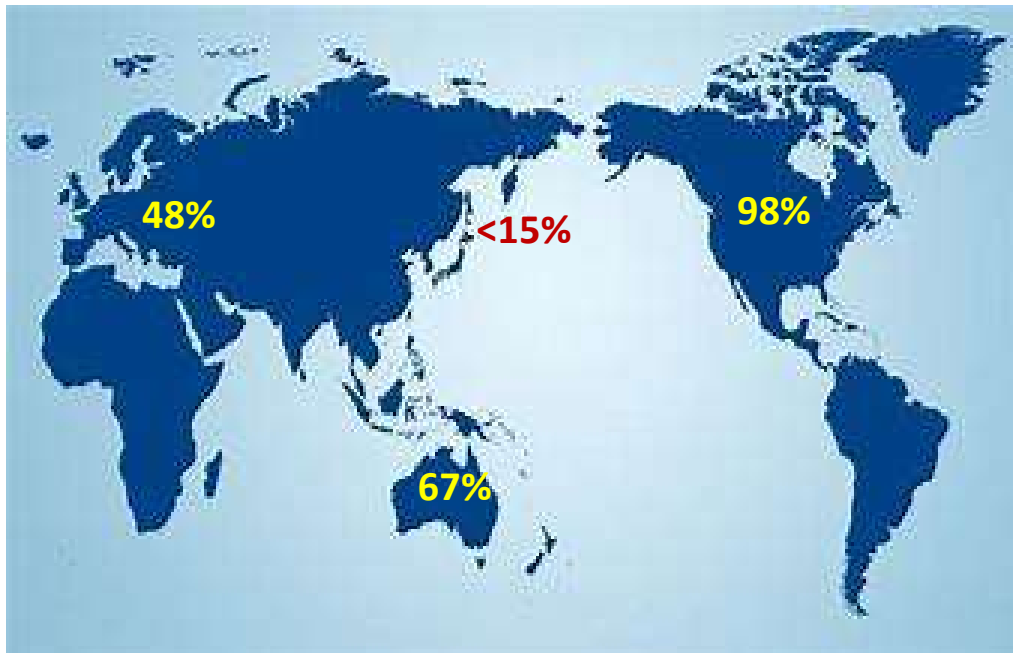
# Benefits of Rapid On-site Cytology Evaluation (ROSE)

Non-  
randomized  
data

| Author                                     | Number | Diagnostic yield with vs. without OCP, % |
|--|--------|--|
| Klapman et al., <sup>96</sup> 2003         | 198    | 78 vs. 32, $p=0.001$                     |
| Alsohaibani et al., <sup>97</sup> 2009     | 104    | 77 vs. 53, $p=0.01$                      |
| Iglesias-Garcia et al., <sup>98</sup> 2011 | 182    | 97 vs. 86, $p=0.01$                      |

- *Provides an immediate cytological diagnosis*
- *Guiding the need for further FNA*
- *Optimizing the diagnostic yield*
- *Minimize the need for repeat EUS*

# Practice of ROSE around the world



Availability of ROSE for EUS FNA practice

*“Limited pathology staffing”*  
= 74%

*“Disbelieve in its additive value”*  
= 32%

*“High costs”*  
= 24%

*“Additional procedure time”*  
= 24%

# “ROSE” is COSTLY!!!

Non-inferiority study: FNA with ROSE vs. standard 7 passes

142 patients were randomized:

- cytopathologist arm (n=73) vs. 7 passes arm (n=69).

***Diagnostic yield for definite diagnosis was similar***

- cytopathology guidance=78.1% vs. 7 passes = 78.3%

***Median charge with onsite cytopathology was significantly greater than performing 7 passes***

- **ROSE= \$1058** versus **7-passes = \$375** (P<0.001)

**ROSE service imposes significant cost to the hospital**  
**→ a major factors for high patient-load, busy hospital**

# Multi-centre, randomized US trial:

## ***EUS-FNA with and without ROSE***

---

US, MCT, RCT, All masses, FNA needles (no FNB)

241 patients were randomized to either: ROSE+ (n=121) or ROSE- (n=120)

### ***No significant difference***

- ***diagnostic yield*** (ROSE+ 75.2% vs. ROSE- 71.6%, P=0.45)
- proportion of inadequate specimens (9.8 vs. 13.3%, P=0.31).
- cytologic characteristics of cellularity, bloodiness, number of cells/slide, and contamination
- procedure time
- adverse events
- number of repeat procedures and costs

Fewer EUS-FNA passes in ROSE+ group (4 vs. 7, P<0.0001).

# Meta-analysis: FNA +/- ROSE

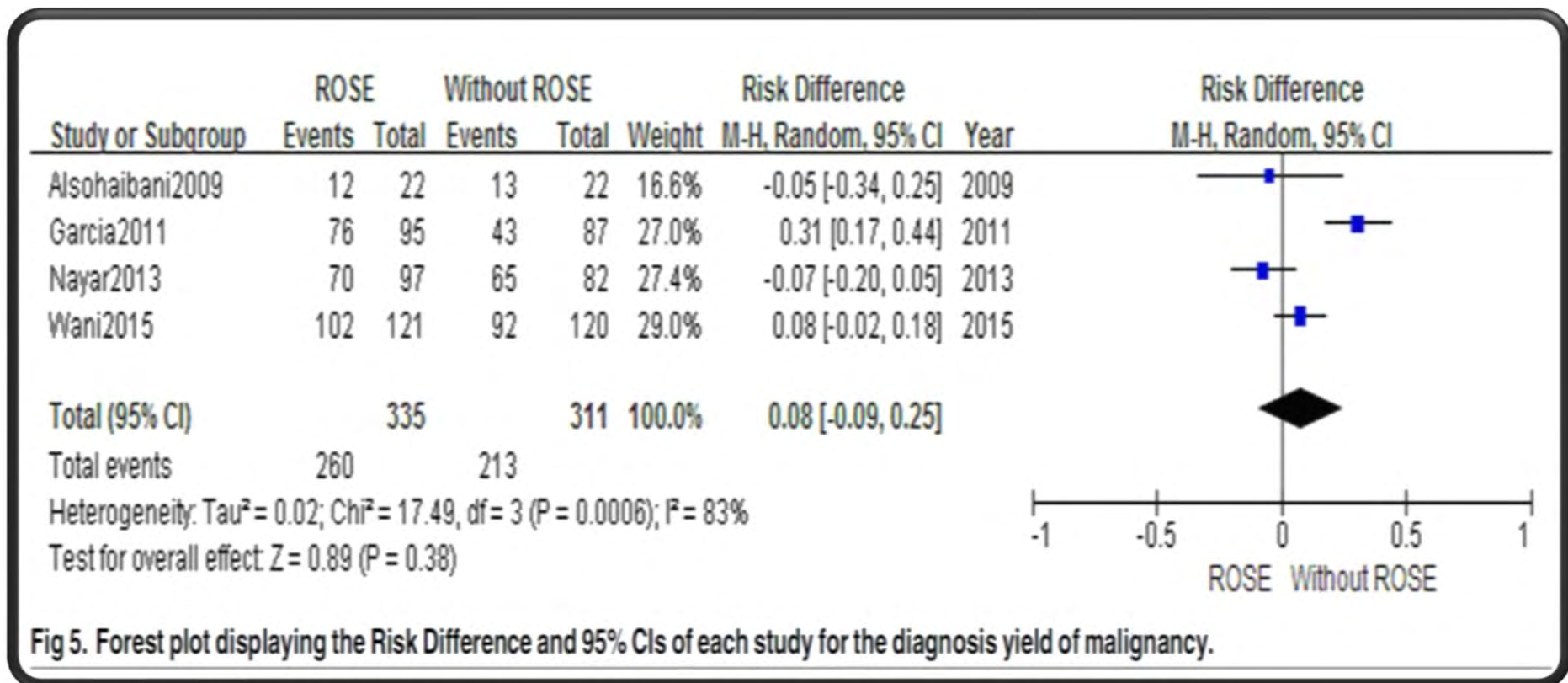


Fig 5. Forest plot displaying the Risk Difference and 95% CIs of each study for the diagnosis yield of malignancy.

# Other weaknesses of ROSE and cytological assessment

- *No ability to differentiate between in-situ vs. invasive cancer*
- *Limited ability to specifically diagnosis*
  - Benign lesions
  - Inflammatory condition
  - Lymphoma
  - Sarcoma/stromal tumour
- *Limited ancillary testing and tumour profiling*
  - ❑ *Halt the progression of “Personalized Oncology therapy”*

## Technical aspects of endoscopic ultrasound (EUS)-guided sampling in gastroenterology: European Society of Gastrointestinal Endoscopy (ESGE) Technical Guideline – March 2017

### On-site cytologic evaluation

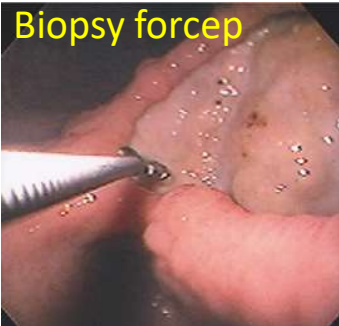
#### RECOMMENDATION

ESGE equally recommends EUS-guided sampling with or without on-site cytologic evaluation (moderate quality evidence, strong recommendation).

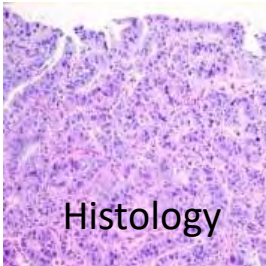
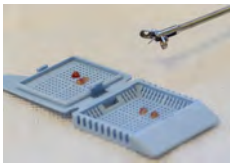
# How can we overcome the lack of ROSE?

## Routine practice of tissue acquisition in luminal Gastroenterology

Biopsy forcep



No ROSE service


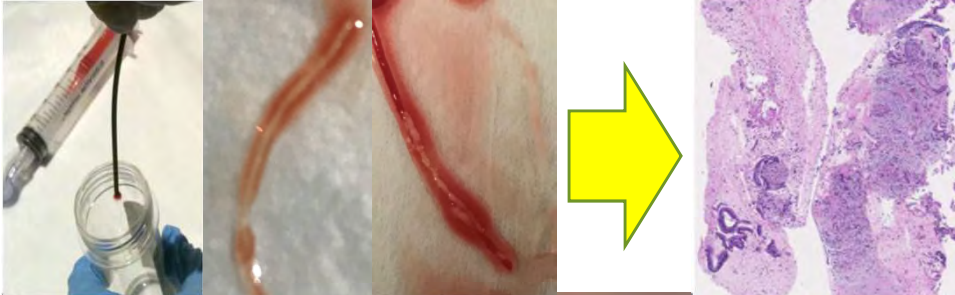



Histology

Tissue diagnosis achieves in >95%

## Ideal trans-luminal tissue acquisition via EUS approach

Flexible core needle

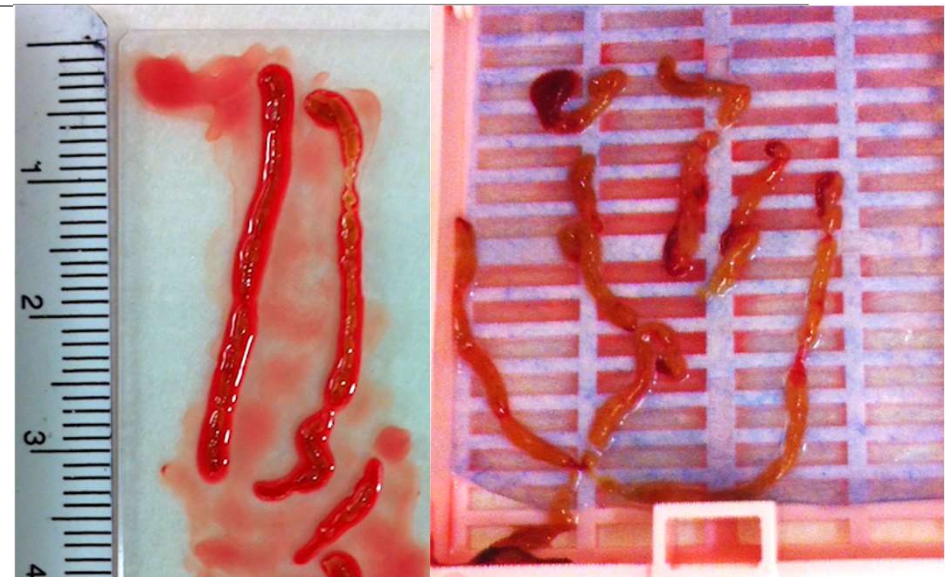
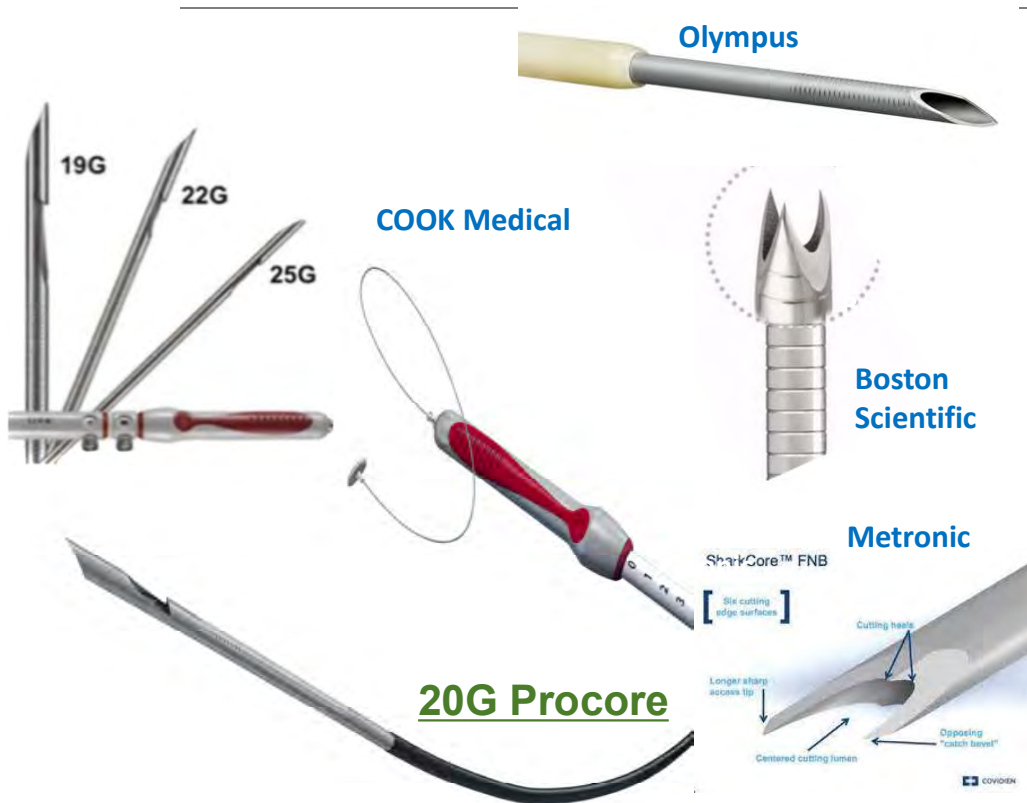


MORE TISSUE



# Advances in EUS needle technology

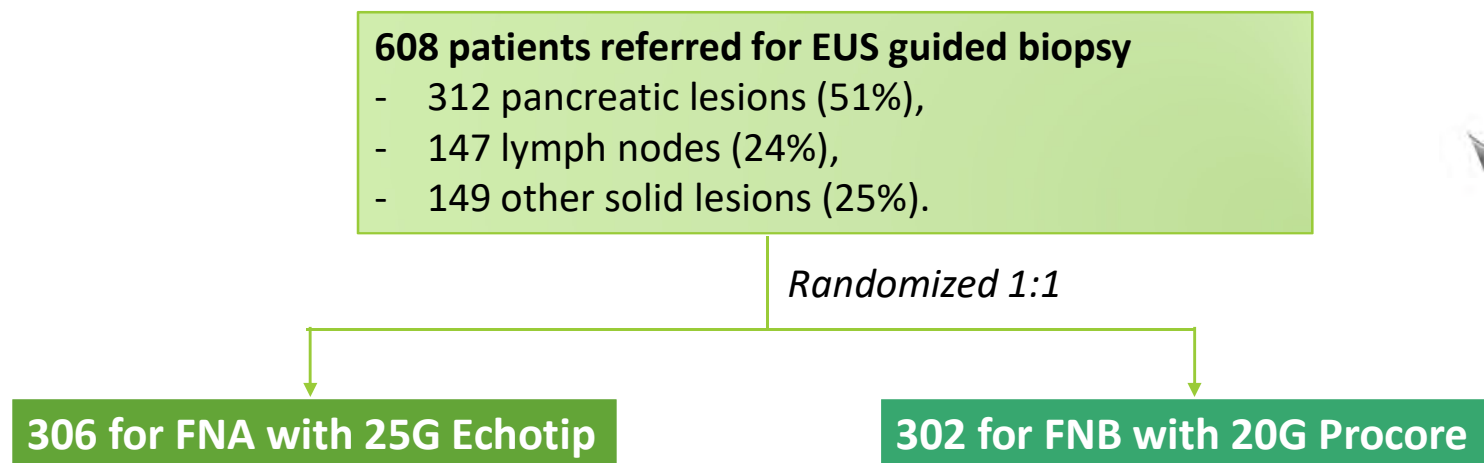
## *...an ideal EUS core needle...*



1. **Core biopsy without ROSE:**
2. **Maximize specificity and accuracy (>90%)**
  - Improve inter-observer variation
3. **Allow routine biomarkers and genomic analyses**
  - Critical for current oncological care of GI cancers
4. **Easy to use, reproducible and safe**

## “ASPRO study”

# MULTICENTER INTERNATIONAL RANDOMIZED TRIAL COMPARING A 25G EUS FINE NEEDLE ASPIRATION DEVICE WITH A 20G EUS FNB DEVICE



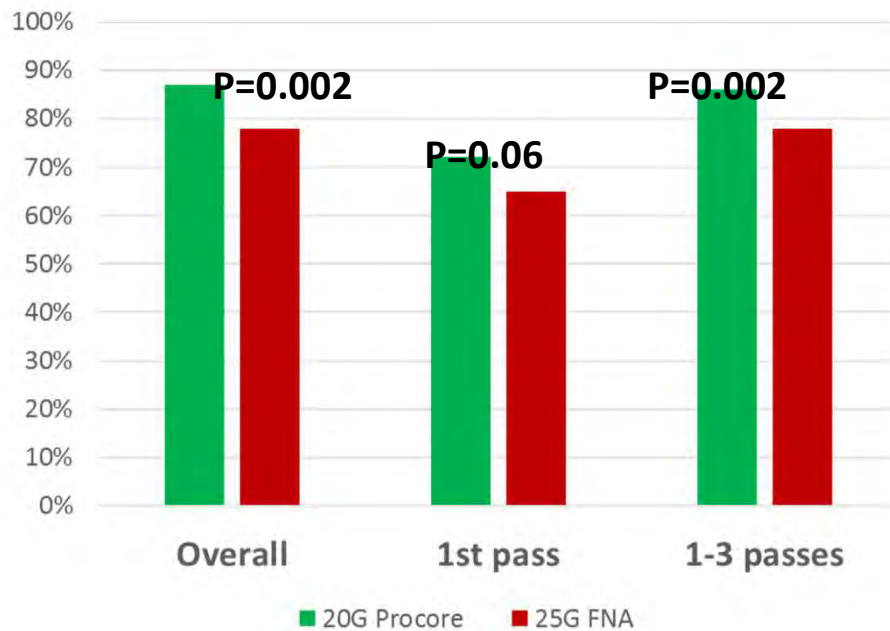
### **Measured outcomes:**

**Primary:** diagnostic accuracy for malignancy and the Bethesda classification (non-diagnostic, benign, atypical, malignant)

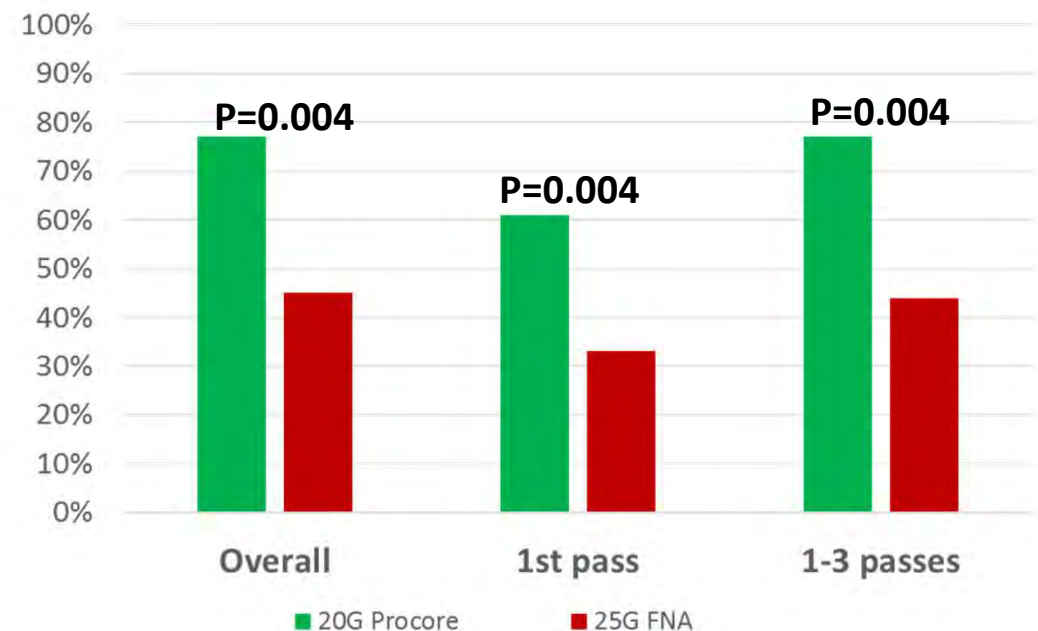
**Secondary:** needle safety, yield per pass, sample sufficiency, cellularity, and histological tissue core yield.

# "ASPRO STUDY" Performance of the new 20G Procore needle

## DIAGNOSTIC ACCURACY



## PRESENCE (%) OF TISSUE CORE

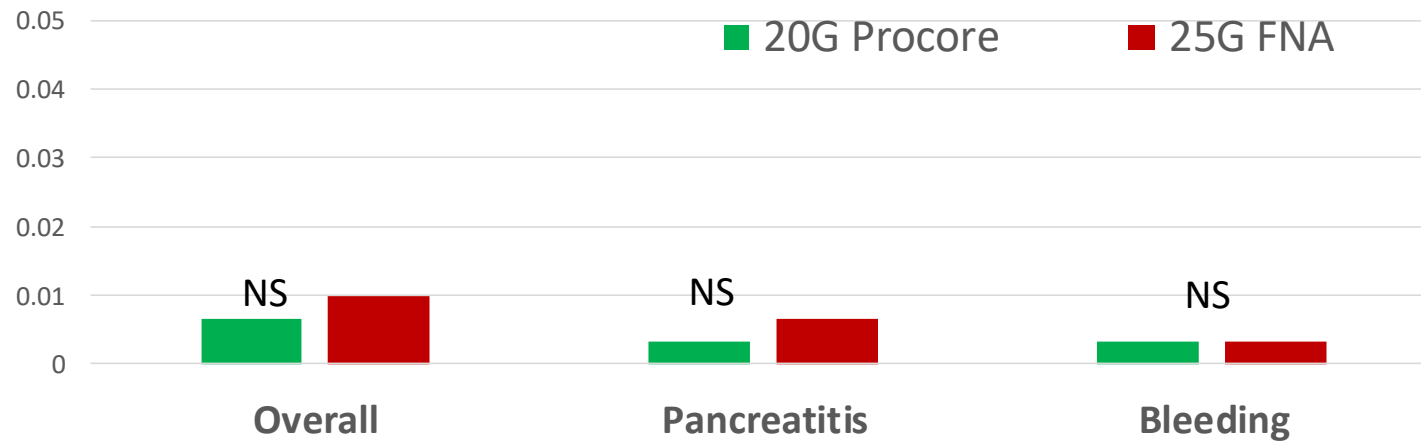


*“ASPRO STUDY”*

# Technical success and complications

|                            | 20G Procore<br>(n=302) | 25G FNA<br>(n=306) |       |
|----------------------------|------------------------|--------------------|-------|
| Technical success rate (%) | 298 (99%)              | 306 (100%)         | 0.043 |

**Complication is Rare = 5/608 (0.8%)**



# Lack of ROSE did not alter diagnostic yield when FNB needles are used!

|                            | Correct diagnosis, % (n/n) | Odds ratio (95% CI) | P-value |
|----------------------------|----------------------------|---------------------|---------|
| <b>Application of ROSE</b> |                            |                     |         |
| Yes                        | 79% (237/299)              | 0.97 (0.51-1.76)    | 0.917   |
| No                         | 85% (232/273)              | *                   |         |

van Riet, Larghi, Nguyen et al. GIE2018

| Subgroup                    | Diagnostic yield (95% CI, I <sup>2</sup> ) |                        |                        |
|-----------------------------|--|------------------------|------------------------|
|                             | All  | Franseen-tip           | Fork-tip               |
| ROSE                        |  |                        |                        |
| Yes                         | 93.7 (87.4-97.0, 77.4)                     | 96.2 (88.1-98.9, 51.5) | 91.2 (79.4-96.6, 71.7) |
| No                          | 95.9 (88.0-98.7, 7.3)                      | 94.1 (80.5-98.4, 0.0)  | 98.8 (88.1-99.9, 0.0)  |
| Statistical difference (P)  | 0.25                                       | 0.26                   | 0.26                   |
| Needle pass                 |  |                        |                        |
| ≤2                          | 90.6 (78.1-96.3, 75.9)                     | 89.5 (66.1-97.4, 80.6) | 93.9 (74.4-98.8, 60.8) |
| >2                          | 93.3 (88.5-96.2, 85.7)                     | 94.4 (85.5-97.9, 91.1) | 91.2 (83.5-95.5, 74.9) |
| Statistical difference, (P) | 0.54                                       | 0.46                   | 0.68                   |

Mohan et al. Endosc Ultrasound 2019

# *FNB (Procore) needles*

## **ROSE did not improve diagnostic yield**

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*Retrospective* from Italian centres

333 patients with pancreatic solid mass lesions

140 cases with ROSE v.s 193 cases without ROSE.

No difference in tissue adequacy

- 92.1 % (ROSE) vs. 88.1 % (no ROSE) (p = 0.227).

***No difference between the ROSE vs. no ROSE in:***

- Sensitivity = 90.7% vs. 87.2%
- Specificity = 100% vs. 100%
- **accuracy = 92.1 % vs. 88.1 %**

No difference in acquire tissue core

- 61.4% (ROSE) vs. 53.4 % (no ROSE) (p = 0.143).

# Ongoing MCT-RCT trials: *FNB and ROSE*

## **FROSENR study (leading site = Larghi)**

A Multicenter Randomized Trial, Comparing EUS Fine Needle Biopsy (EUS-FNB) with Rapid on-Site Evaluation (ROSE) versus EUS-FNB Alone for the evaluation of Patients with Solid Pancreatic Lesions.

Sample size = 730;

16 sites (Europe, Asia, Australia)

1:1 randomization between FNB/ROSE vs. FNB alone

*Crino et al. Dig Liver Dis. 2019*

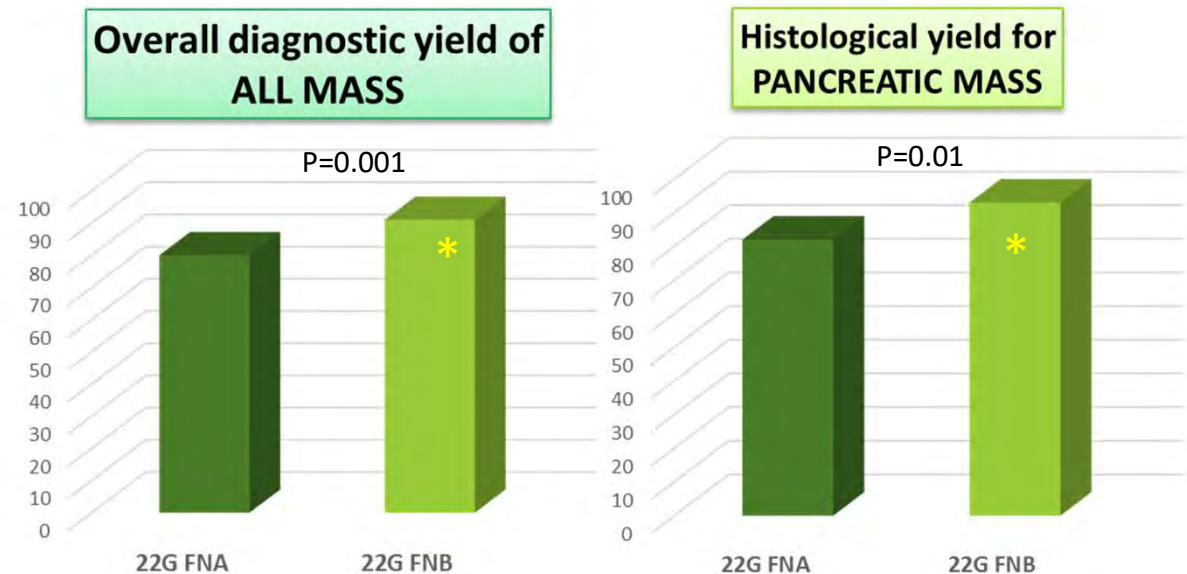
## **BEAT-ROSE study (leading site = Nguyen)**

*Evaluation of fine needle biopsy (FNB) for EUS guided tissue acquisition of pancreatic masses to negate the need for rapid on-site evaluation: a multi-centre randomized control trial*

- ▶ Sample size = 598;
- ▶ 12 sites in Australian and Asia-Pacific region
- ▶ 1:1 randomization between FNA/ROSE vs. FNB alone

# FNB gives higher diagnostic and histological yield for pancreatic mass

- RCT of 5 centres
- n = 408; solid mass (>1 cm), 4 passes each patients
- No ROSE
- Randomized to:
  - FNA (22G, n = 190)or
  - FNB (22G Procore n = 187)



No difference in diagnostic yield between EUS-FNA and EUS-FNB for *non-pancreatic masses*.



# Fork tip needle



- **Multicenter retrospective study**
  - 147 patients: 101 EUS-FNB (Fork-tip) vs. 46 EUS-FNA needles.
  - All solid masses
  - 80% without ROSE
- **Compared to FNA needle, Fork-tip FNB needle had higher:**
  - Diagnostic yield (89% vs 37% (P = 0.001))
  - Cytopathology yield (92% vs 46% (P = 0.001))
  - IHC yield (89% vs 41% (P = 0.001))
- **Multicenter prospective study**
  - 100
  - 3 centers
  - Solid pancreatic masses (28.5 mm (SD 11.7))
  - Diagnostic accuracy was 93%.
  - Core specimens in 67% of patients
    - poor agreement with MOSE (kappa, 0.12; 95% CI: 0.03-0.28).

# Franseen tip needle



- Prospective multi-centre study (n=200; FNB of solid lesions with ROSE).
- Tissue obtained by EUS-FNB was adequate for evaluation and **diagnosis by ROSE in 197/200 cases (98.5%)**.
- *Core of tissue was obtained in 131/145 (90%) of cases*

*Adler et al. Endosc Ultrasound 2019*

## Prospective tandem study:

- N=56; pancreatic solid mass
- First pass = 22G Franseen needle
- Second pass = 22G FNA needle
- Formalin fixed for histology
- No ROSE

| No ROSE     | Franseen needle<br>(n=56), n (%) | Standard needle<br>(n=56), n (%) | P     |
|-------------|----------------------------------|----------------------------------|-------|
| Sensitivity | 42/52 (80.7)                     | 31/52 (59.6)                     | 0.018 |
| Specificity | 4/4 (100)                        | 4/4 (100)                        | NS    |
| PPV         | 42/42 (100)                      | 31/31 (100)                      | NS    |
| NPV         | 4/14 (28.6)                      | 4/25 (16.0)                      | 0.351 |
| Accuracy    | 44/52 (84.6)                     | 33/52 (63.5)                     | 0.014 |

*Matsuno et al. Endosc Ultrasound 2019*

# Randomized comparison: *Franseen vs. standard FNA needles*

- N= 46, solid lesion (mainly pancreatic) randomized to either:
  - 22G Franseen needle
  - 22G Standard needle
  - 2 passes per lesion, presence of ROSE

|                              | Franseen (46)        | Standard (46)        | p-value    |
|------------------------------|----------------------|----------------------|------------|
| <b>Diagnostic Cell Block</b> | <b>97.8%</b>         | <b>82.6%</b>         | <b>.03</b> |
| Tumor                        | 0.68 mm <sup>2</sup> | .099 mm <sup>2</sup> | .0001      |
| Retained Architect           | 93.5%                | 19.6%                | .0001      |
| Total Tissue                 | 6.1 mm <sup>2</sup>  | 0.28 mm <sup>2</sup> | .0001      |

# 19G Needle and “core” specimen

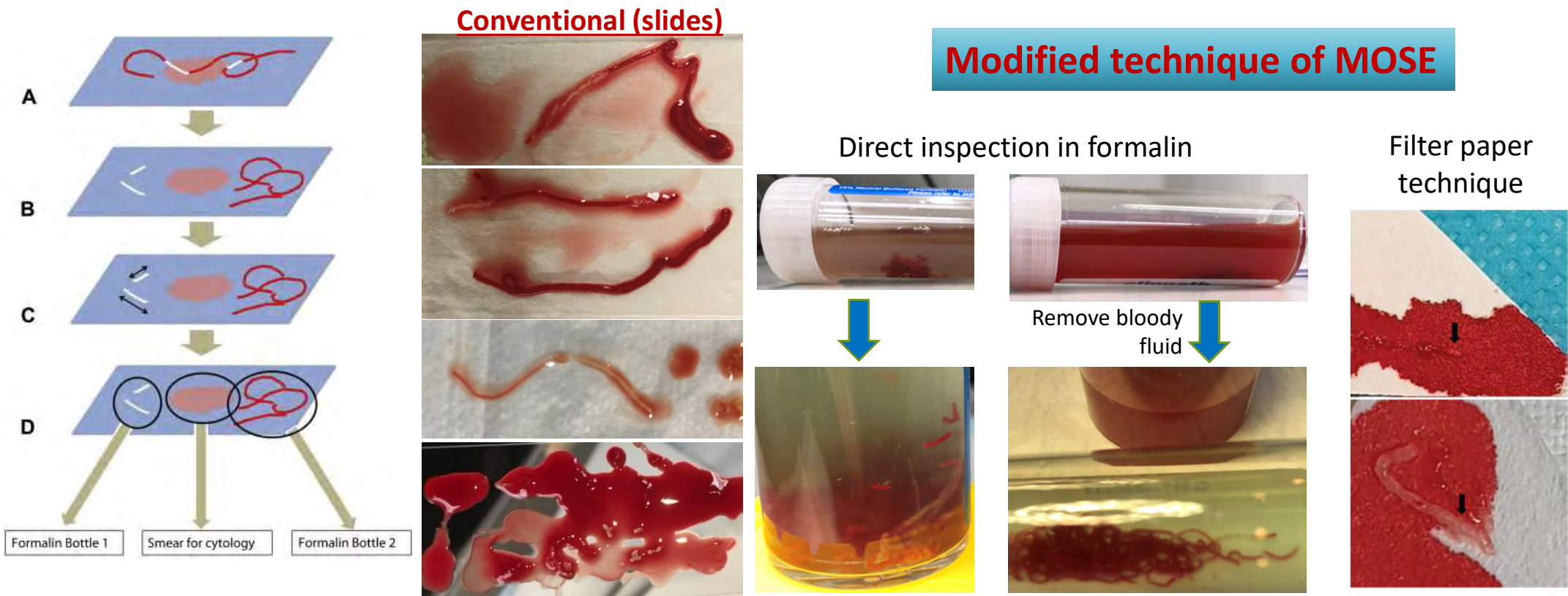


| Study, Year                        | Needle Size (G) | Number of Patients | Histologic Adequacy (%) | Location of Biopsy    |
|------------------------------------|-----------------|--------------------|-------------------------|-----------------------|
| Iwashita et al, <sup>47</sup> 2012 | 19              | 44                 | 43                      | Pancreas <sup>a</sup> |
| Yasuda et al, <sup>45</sup> 2006   | 19              | 104                | 98                      | Lymph nodes           |
| Rong et al. <sup>44</sup> 2012     | 22              | 54                 | 70.4                    | Pancreas              |

**Use of 19G needles is limited by its poor technical success and risk of bleeding/pancreatitis**

|  |                 |     |      |  |
|--|-----------------|-----|------|--|
| Bang et al, <sup>43</sup> 2012         | 22, FNA         | 28  | 66.7 | Pancreas   |
|  | 22, FNB         | 28  | 80   |  |
| Larghi et al, <sup>46</sup> 2011       | 19              | 120 | 97.5 | Various  |
| Varadarajulu et al, <sup>48</sup> 2012 | 19 <sup>b</sup> | 38  | 94.7 | Subepithelial masses<br>Pancreatic (head and uncinata lesions) |

# Macroscopic Onsite Examination (MOSE) as an alternative technique for assessing specimens from FNB needles.



# Direct MOSE to guide the number of pass

- 46 consecutive patients with 54 solid lesions were biopsied using FNB needle (22G Franseen). Retrospective reviewed.
- If no macroscopic core was visualized, a second pass was performed.
- **Core tissue was visualized in 93% (50/54) of targets with a single pass**
  - Histologic core fragments confirmed in 94% (47/50).
- **Overall correlation between MOSE and histologic core fragments was 94% (48/51).**
- Diagnostic adequacy was 98% (53/54) with one biliary target biopsied without significant material.
- The overall diagnostic accuracy was 94 %, with 100% specificity.

# Tissue specimen from FNB improves inter-observer agreement

## 20-gauge ProCore FNB

Diagnostic classification

Sufficient quality in 91%\*

Tissue cores in 70% \*

≥50% target cells 70%

Additional analysis in 76%\*

## 25-gauge Echotopic FNA

Diagnostic classification\*

\*P<0.05; 25G vs. 20G Procore)

Kappa value

*expert vs non-expert*

0.62 = 0.59

0.51 > 0.42

0.41 > 0.26

0.33 = 0.27

0.51 > 0.38

*expert vs non-expert*

0.48 > 0.35

No differences  
between pathologists

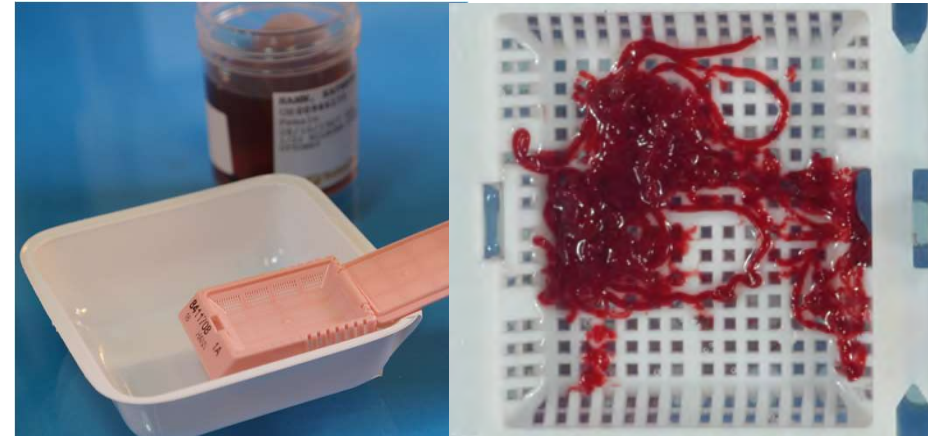
Non-expert pathologists performed less well

**Histology provided better agreement than cytology, but only when a core specimen was obtained with FNB (P = 0.004 vs P = 0.432).**

# RAH approach for EUS FNA/FNB sampling and tissue processing

Predominantly core (FNB) needles with direct histology tissue processing

20G  
Procore  
(at least 2  
passes) \*



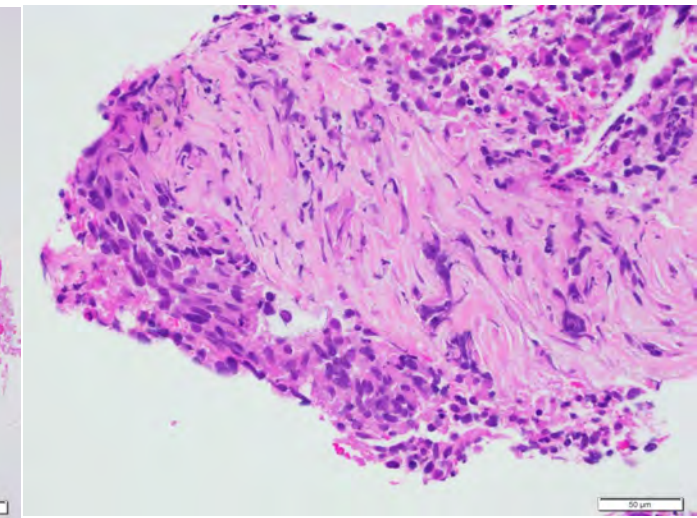
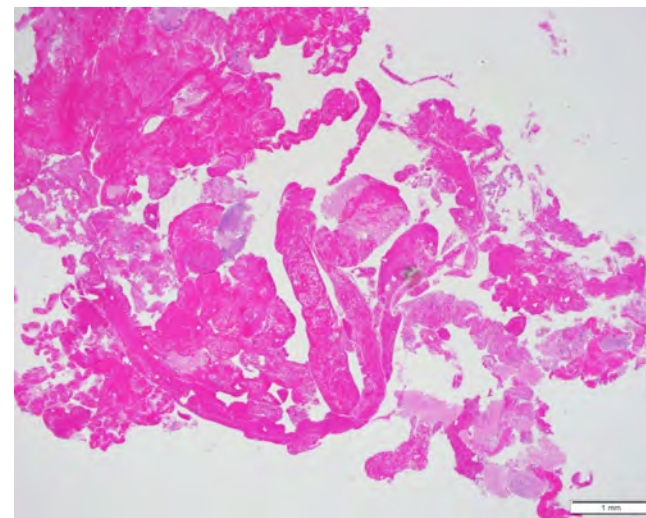
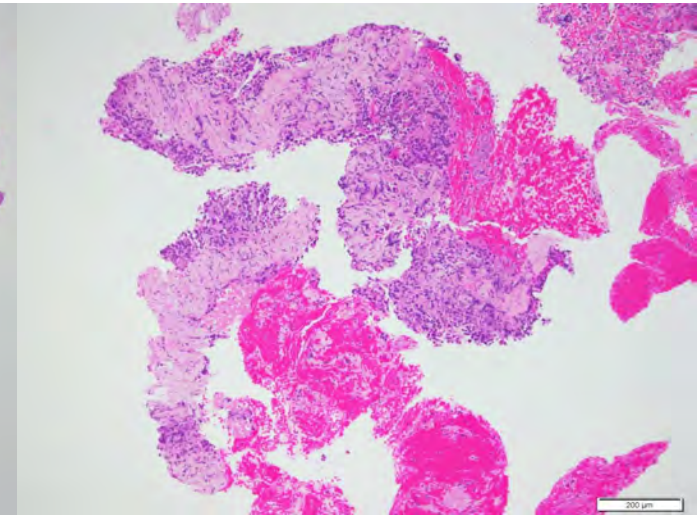
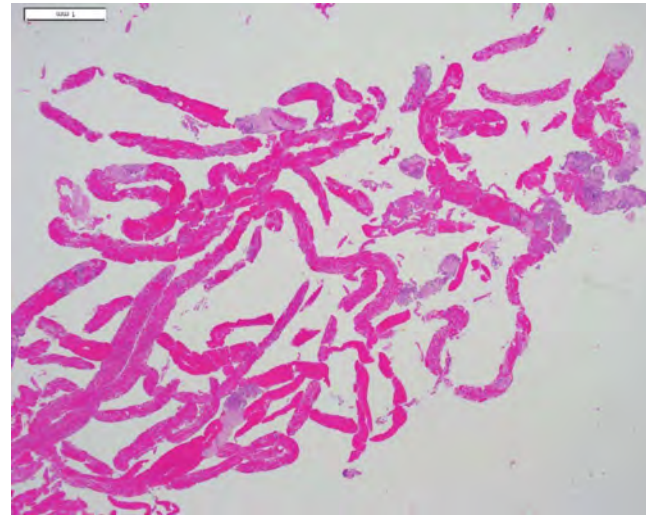
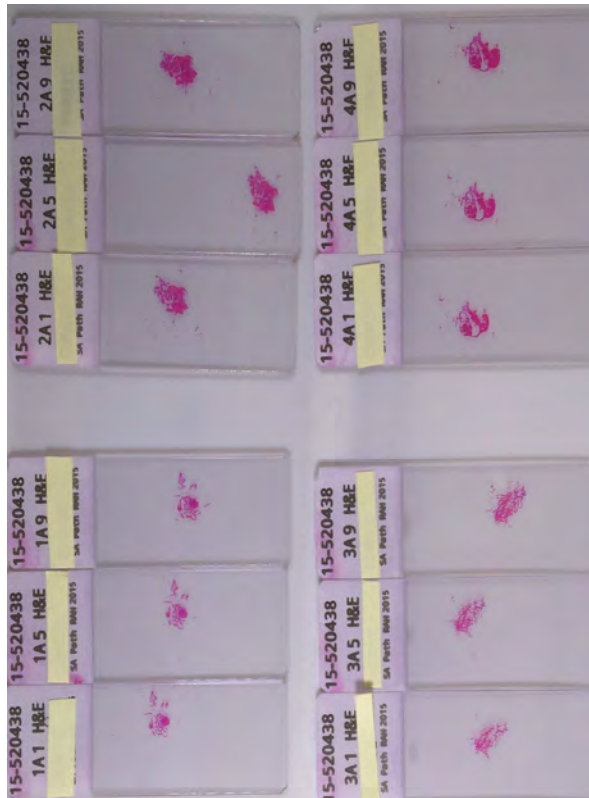
\* If FNA needle use, at least 4 passes.

Extraction of aspirate by flushing the needle with small amount of 0.9% saline solution into fixative solution.

- No smears from aspirate
- No on site assessment by cytologist
- Entire material for paraffin block



**RIBBON OF 12 (1<sup>ST</sup> 5<sup>TH</sup> AND 9<sup>TH</sup> - HE STAIN, SPARES ON "CHARGED" SLIDES**



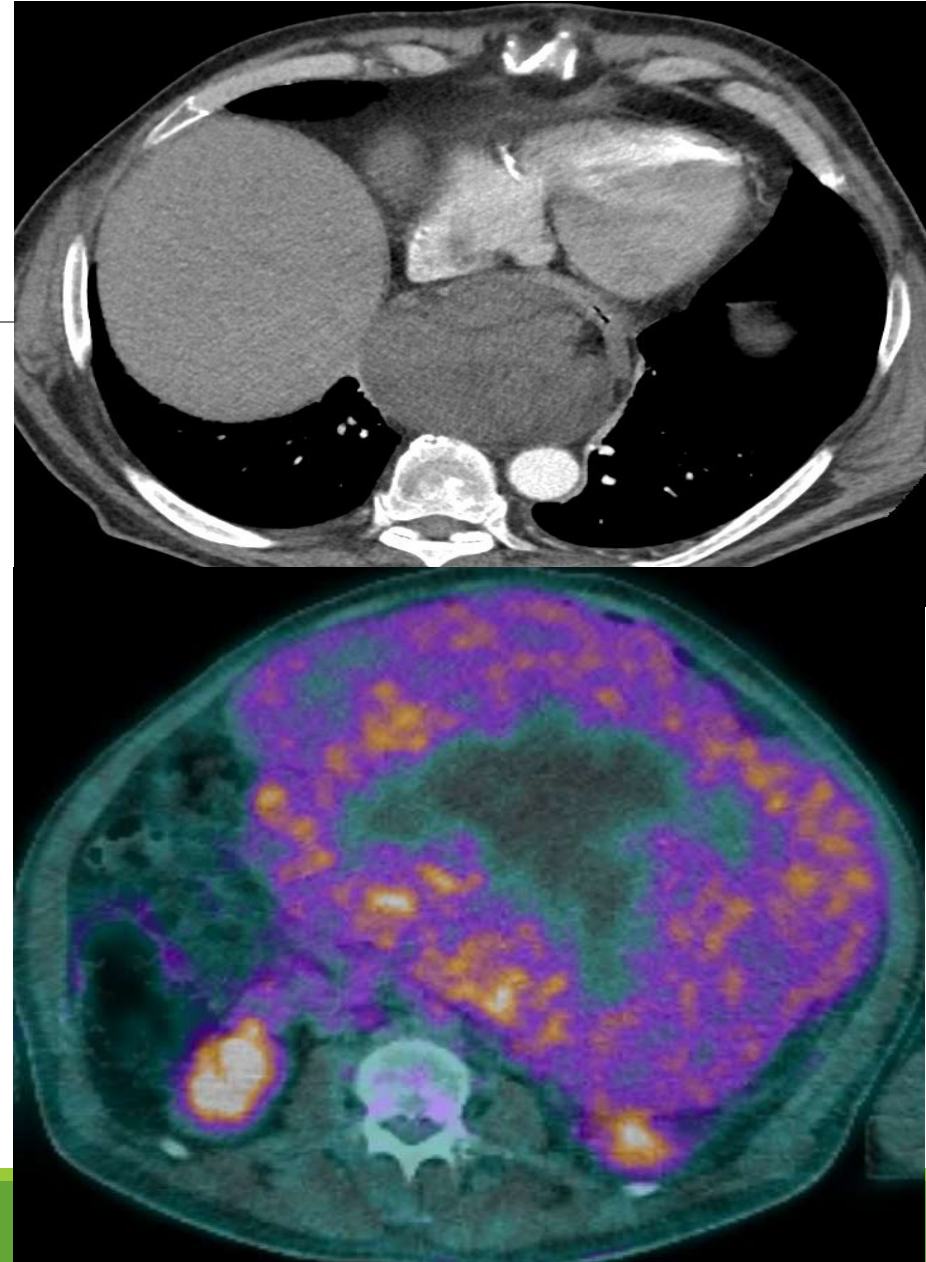
# Case *illustrations*

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# ***Massive abdominal mass...***

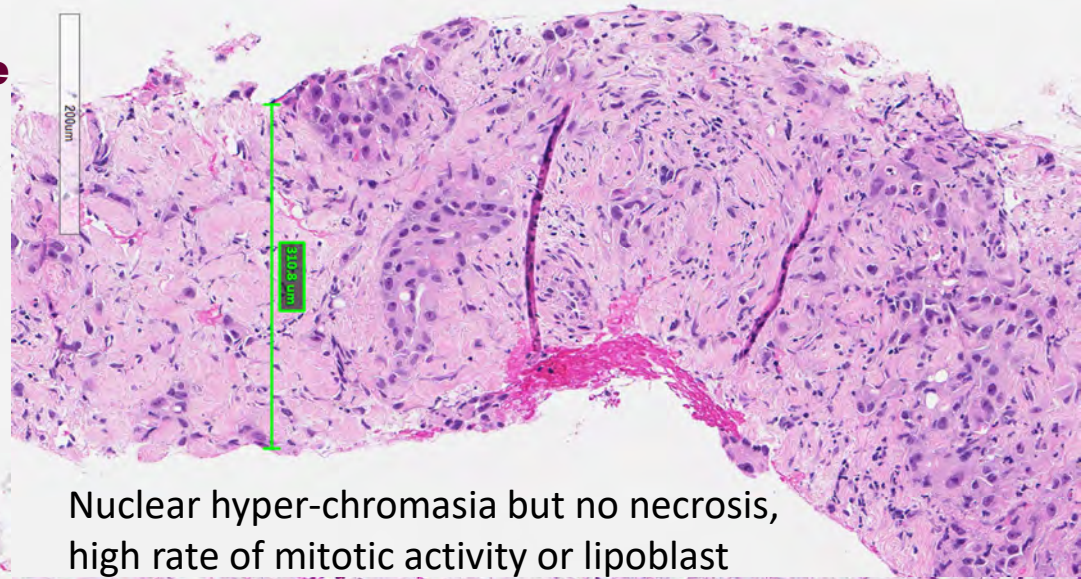
- 67 man
- 6 months of upper abdominal pain
- increasingly abdominal distension
- markedly reduced appetite and oral intake
- CBE, ECaU, LFT - all normal
- Imaging:
  - CT and MRI
  - PET scan



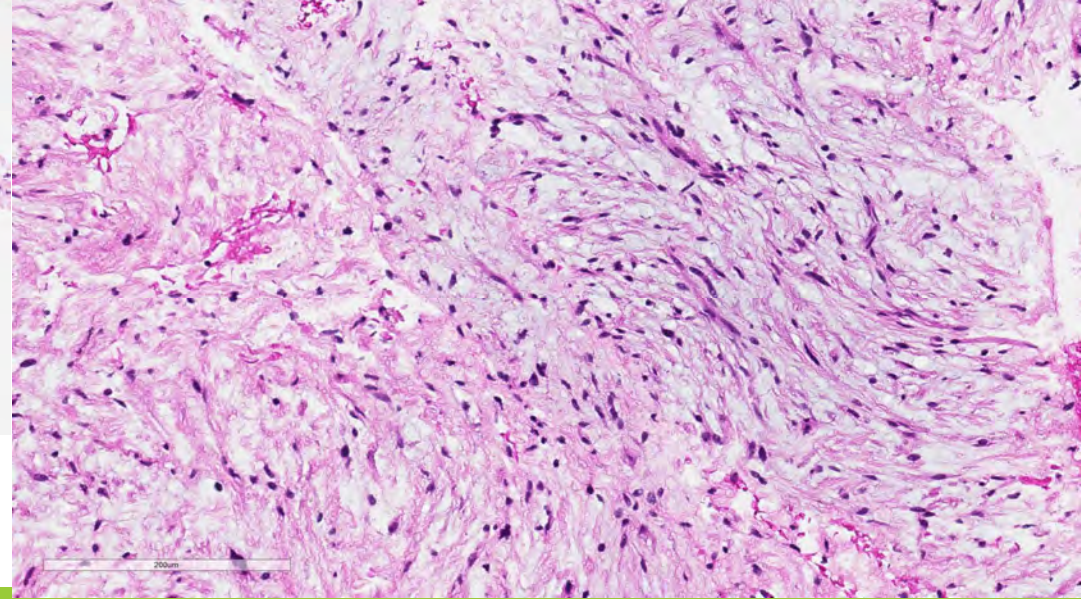
**A single pass of 22G Core needle**



***Cores of monotonous spindle cell proliferation  
-with a myxoid background***



**Nuclear hyper-chromasia but no necrosis,  
high rate of mitotic activity or lipoblast**



# ***Immunohistochemistry and molecular evaluation***

---

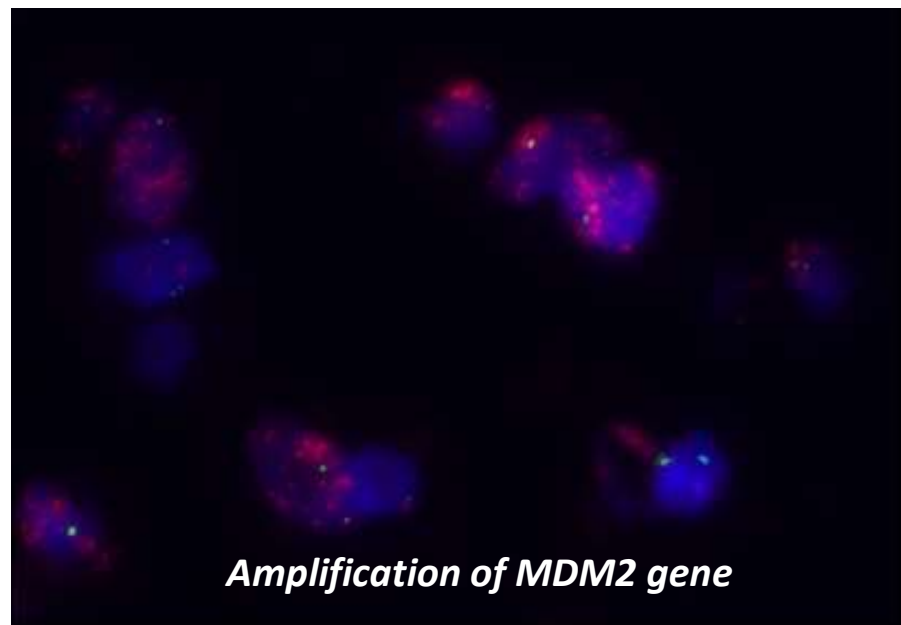
## **Immuno-histochemistry assessment:**

Negative stain for:

- Desmin
- DOG1
- C-kit
- AE1/3

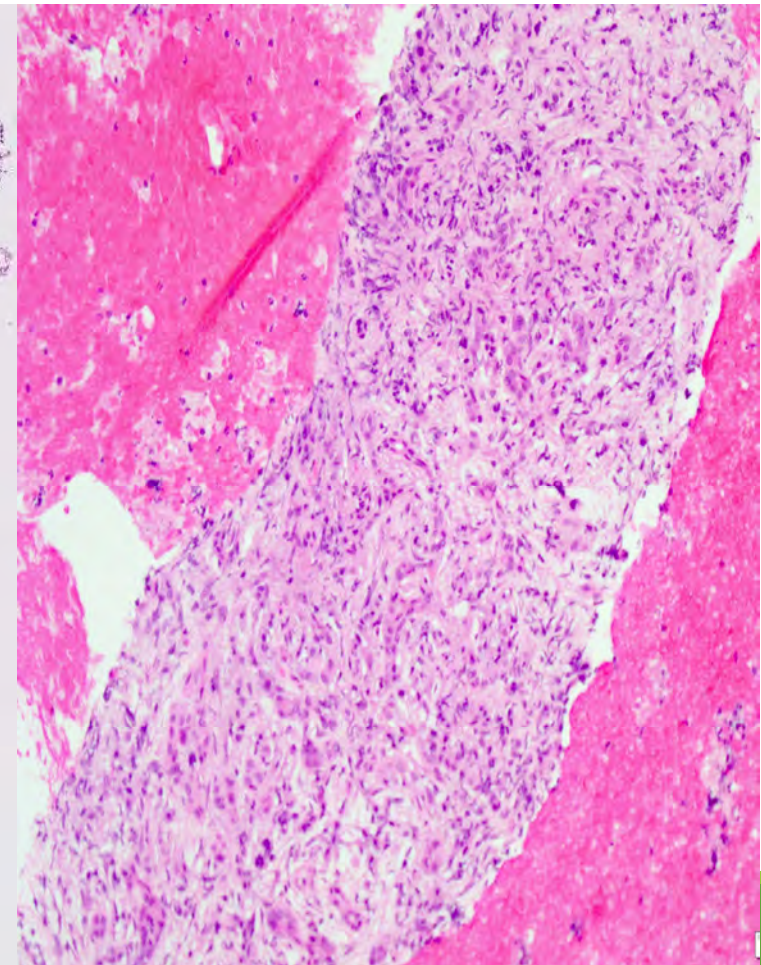
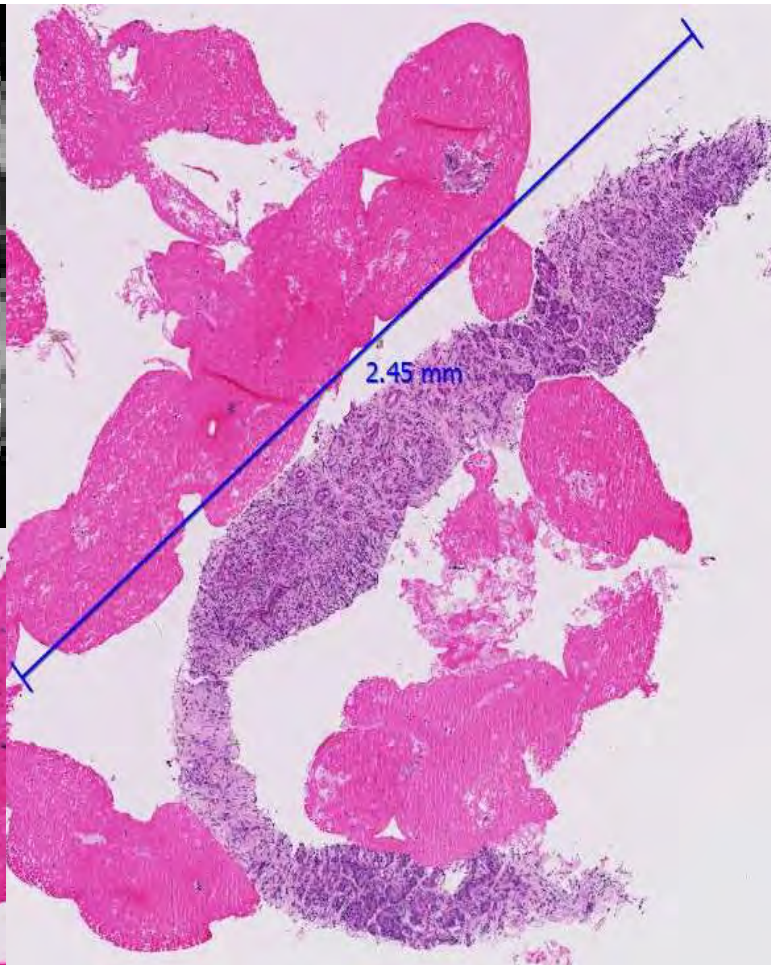
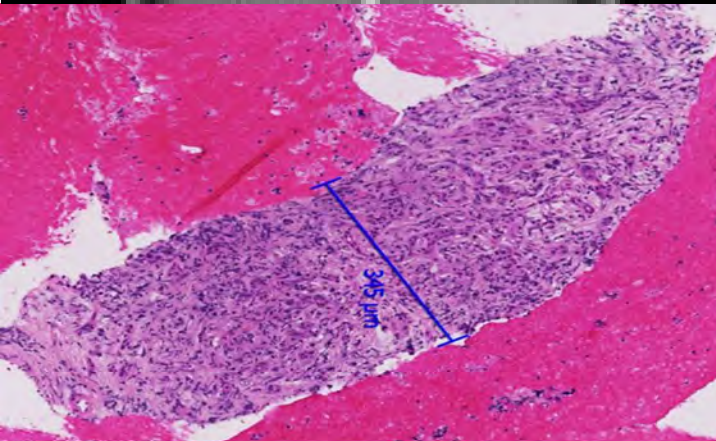
***Not GIST***

## **SISH and FISH analyses:**



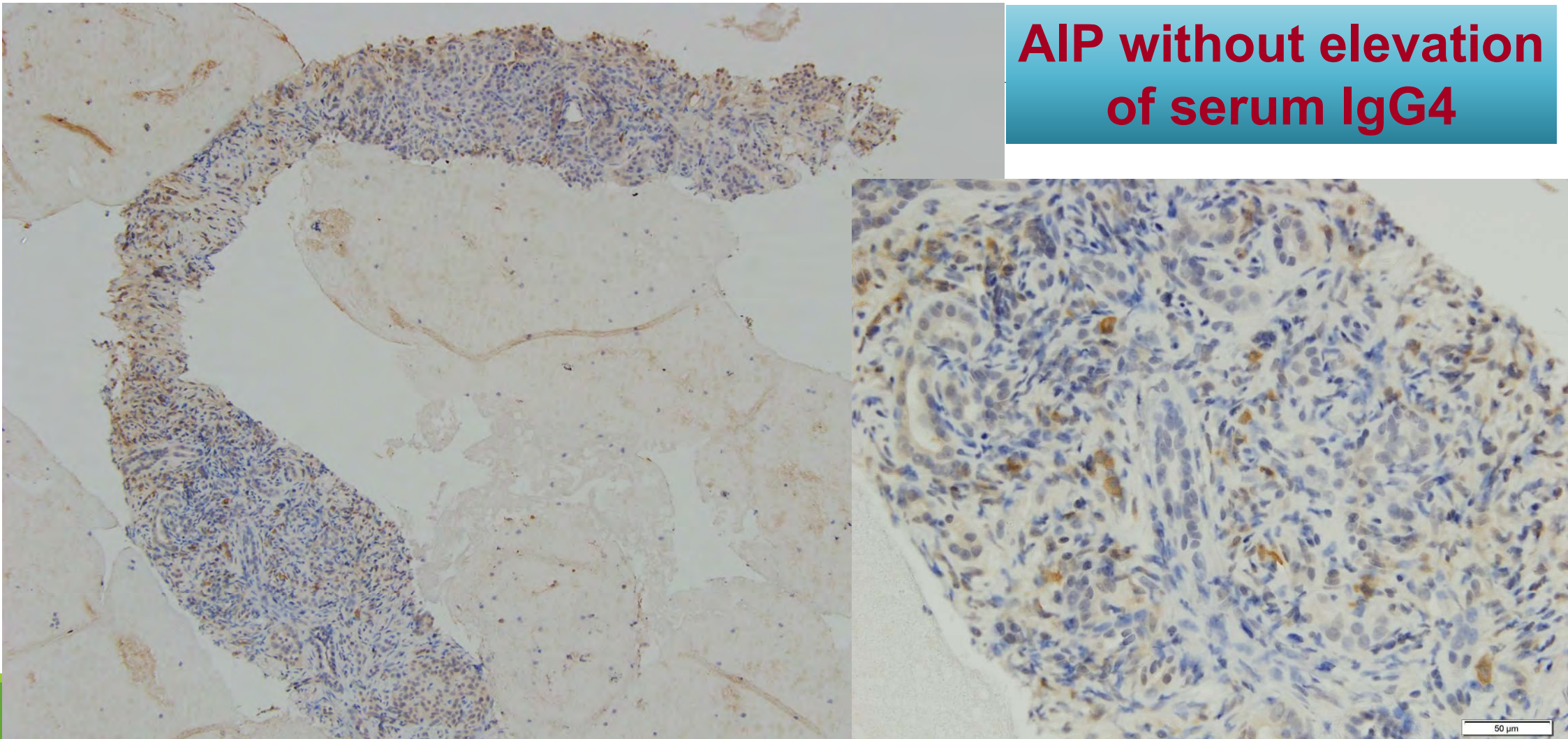
**Dedifferentiated liposarcoma → Surgery, completely resected**

# Jaundice with a “sausage shape” pancreas

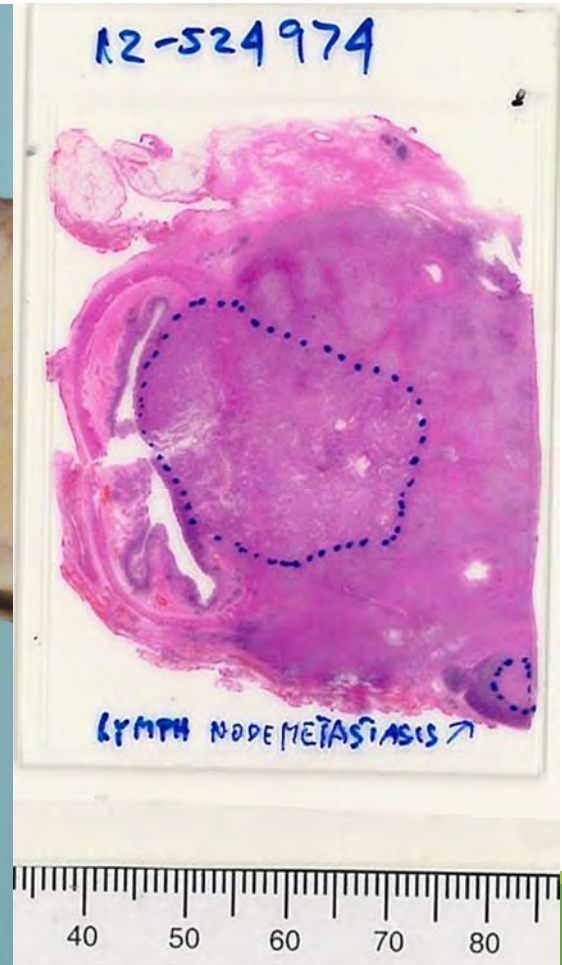
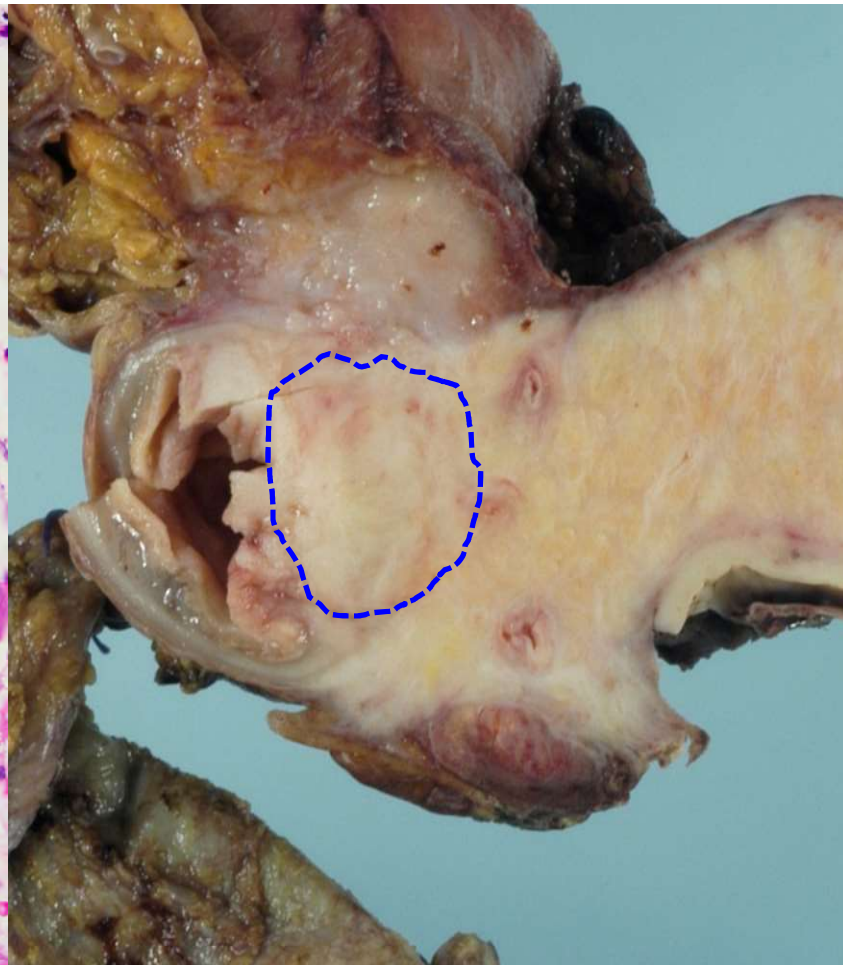
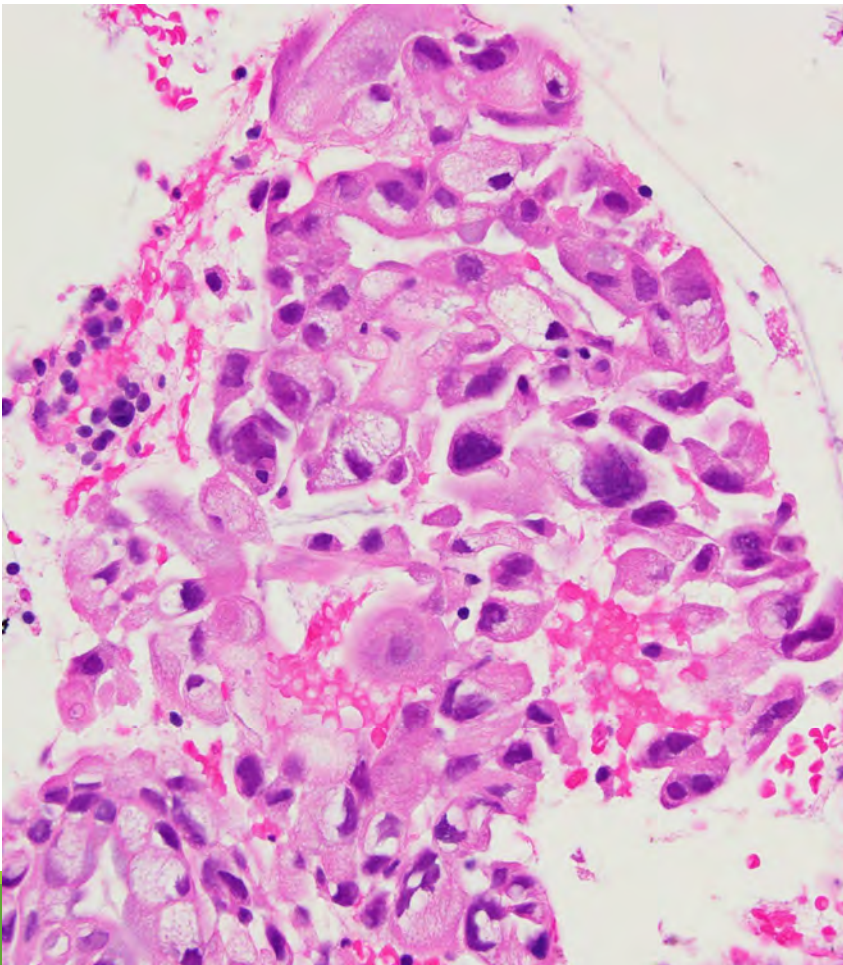


# ***Immunohistochemistry stain***

**AIP without elevation  
of serum IgG4**



# ***A surprising finding of... adenosquamous carcinoma...***

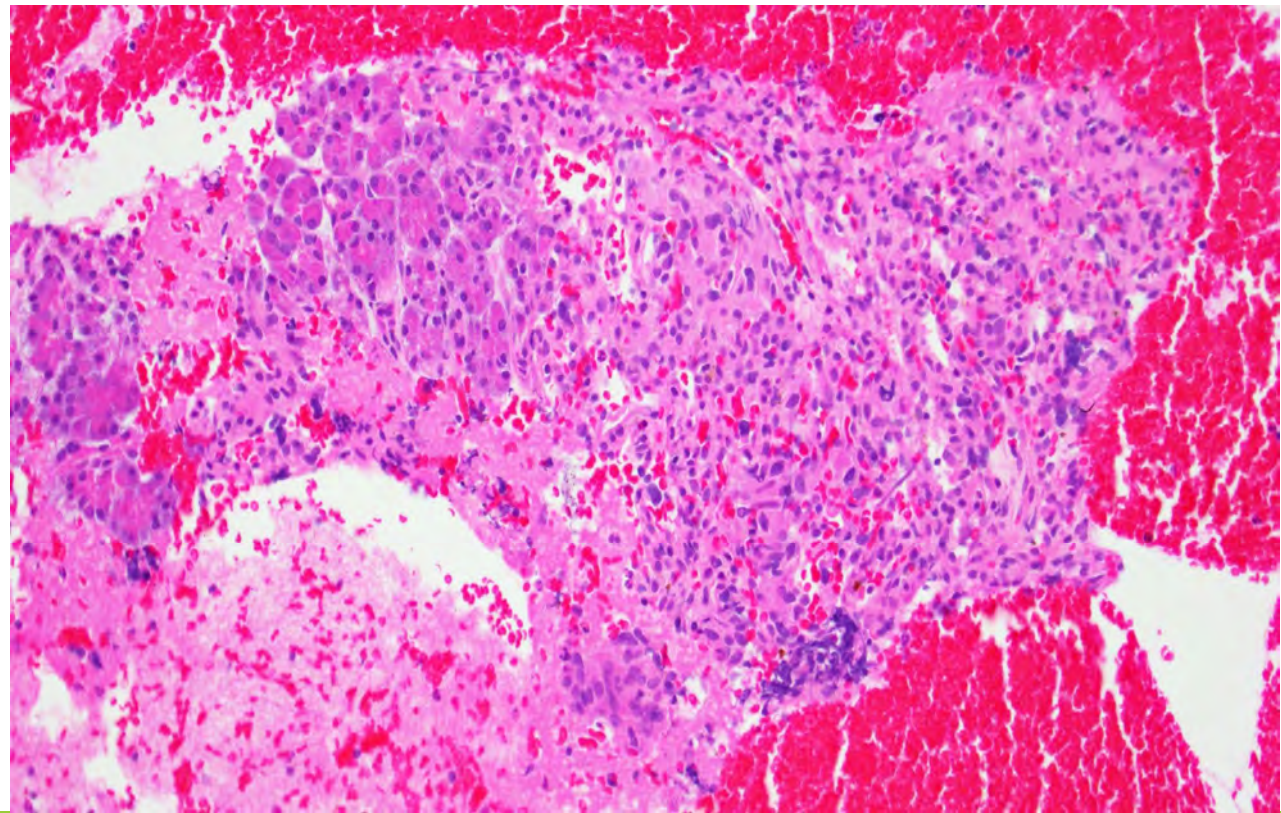


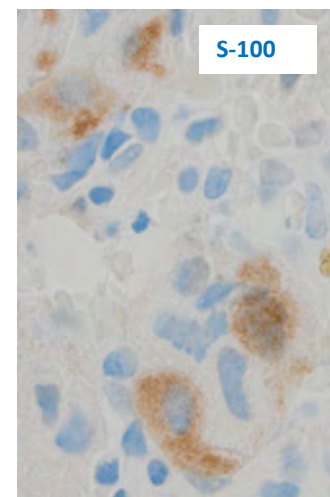
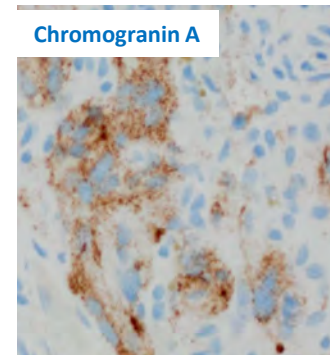
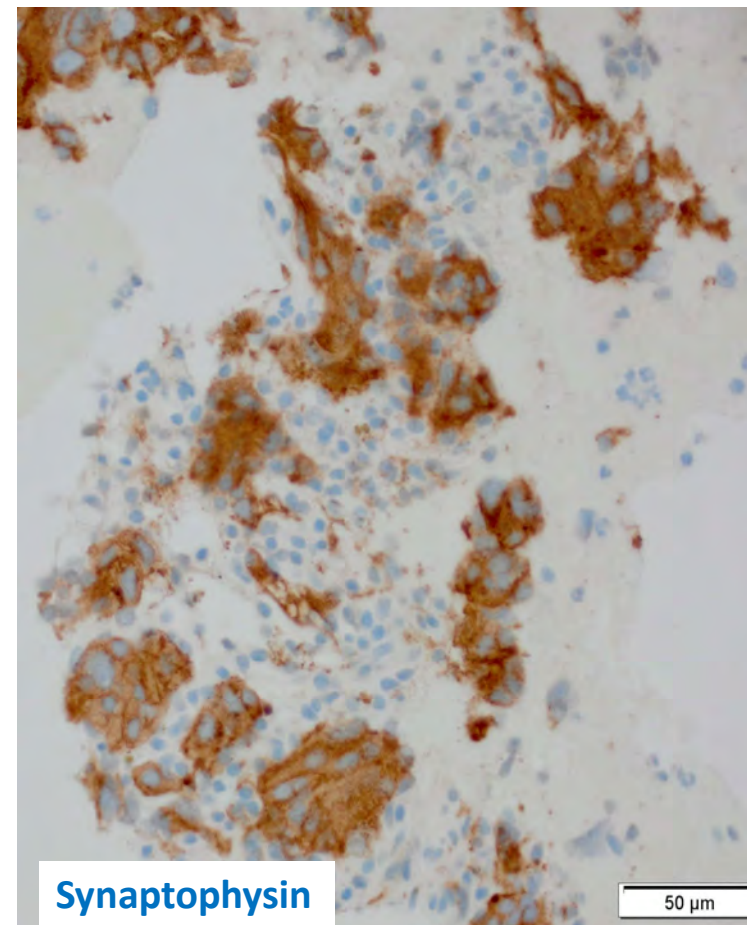
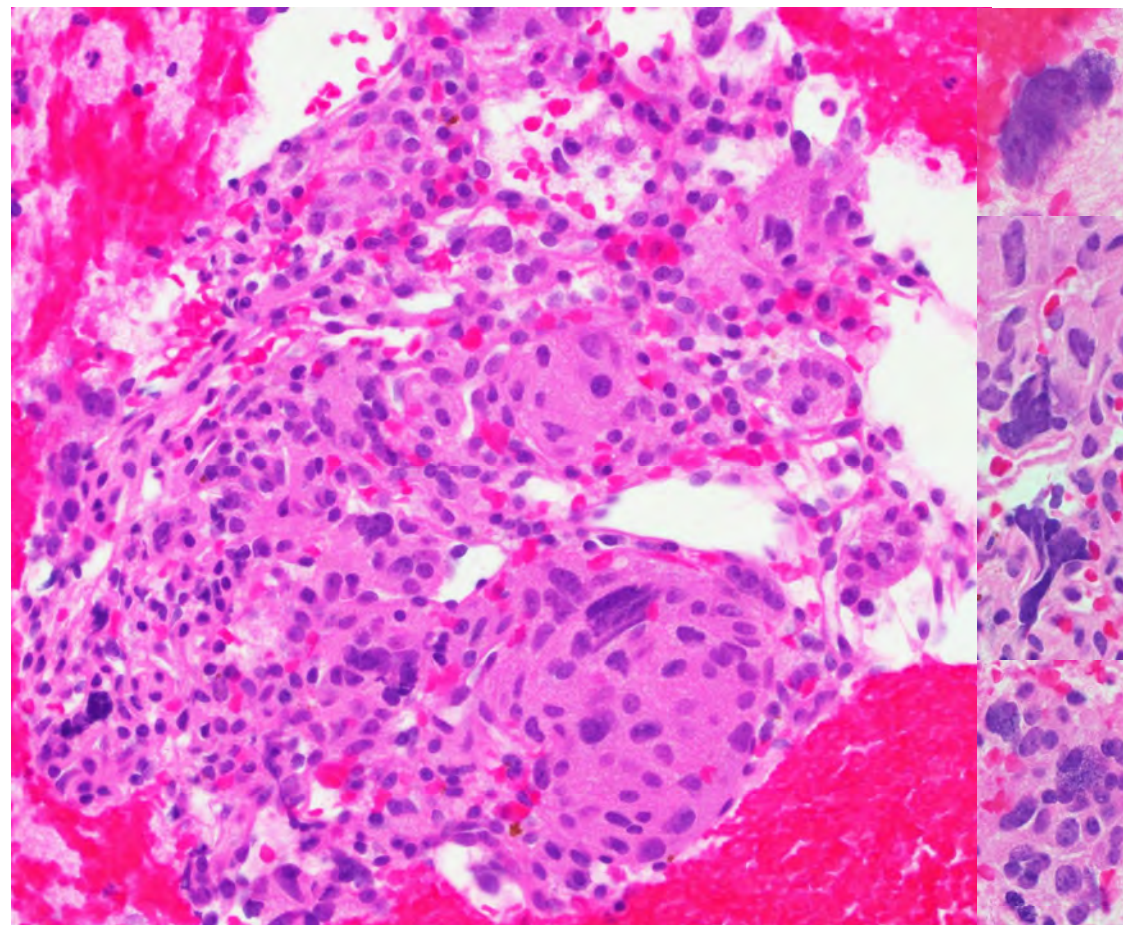


# Direct histology for conventional FNA needle is also useful...

---

- 62/F
- Surgery 7 months ago for ovarian papillary serous carcinoma
- Completed 1<sup>st</sup> chemotherapy course
- 8mm pancreatic head lesion PET +ve
- ? Metastatic deposit
- EUS FNA biopsy (22G).





## Extra-adrenal Paraganglioma

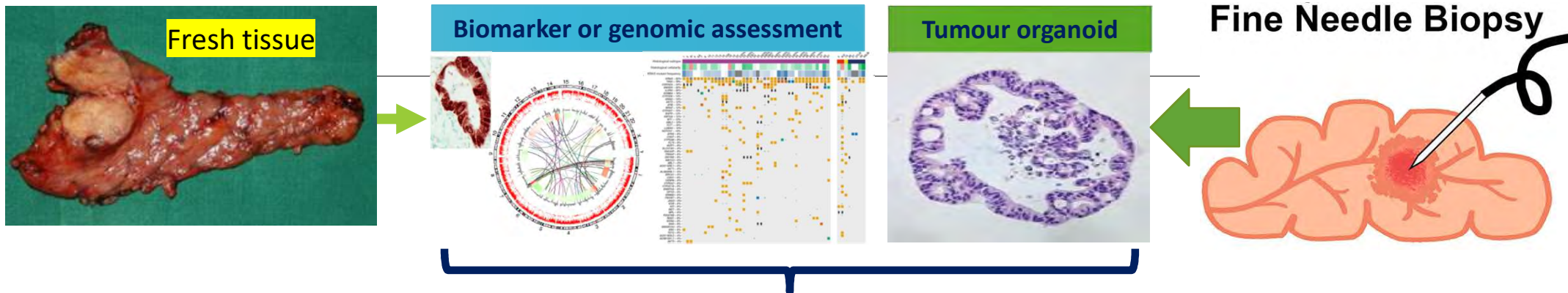
- Rare neuroendocrine neoplasms arising in extra-adrenal chromaffin cells of autonomic nervous system

# **Future implications of EUS guided core biopsy**

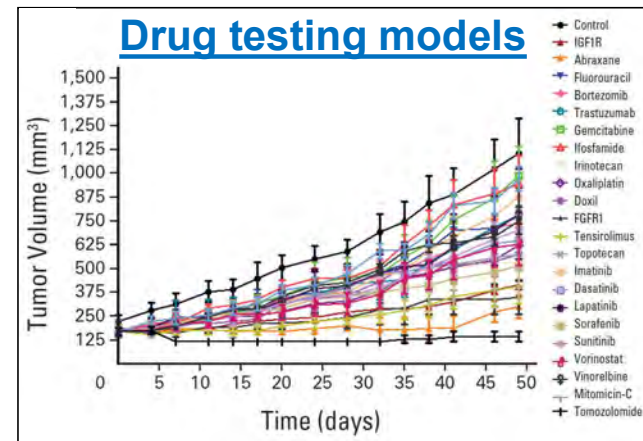
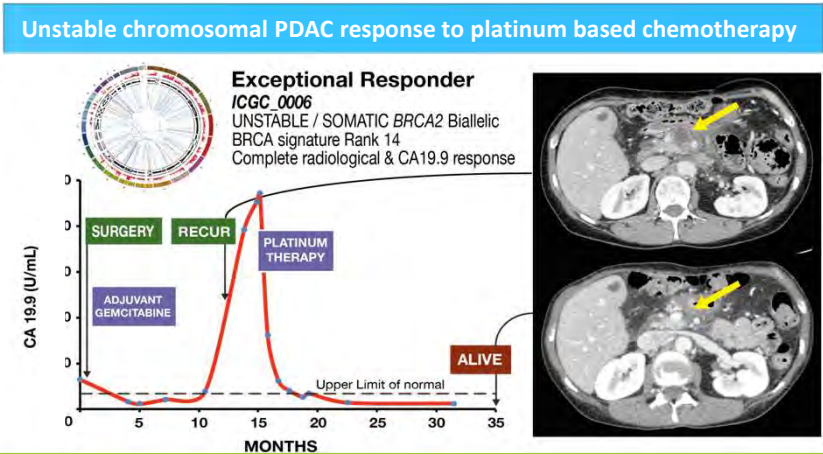
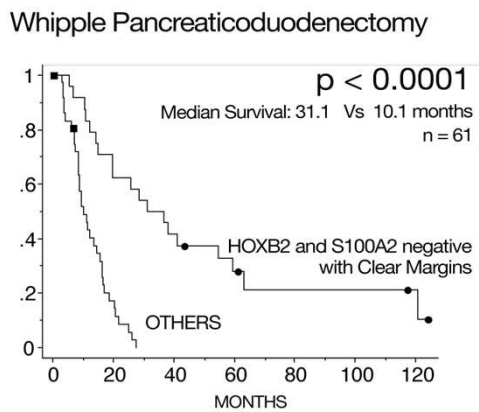
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# Provides tissue for Precision Oncology



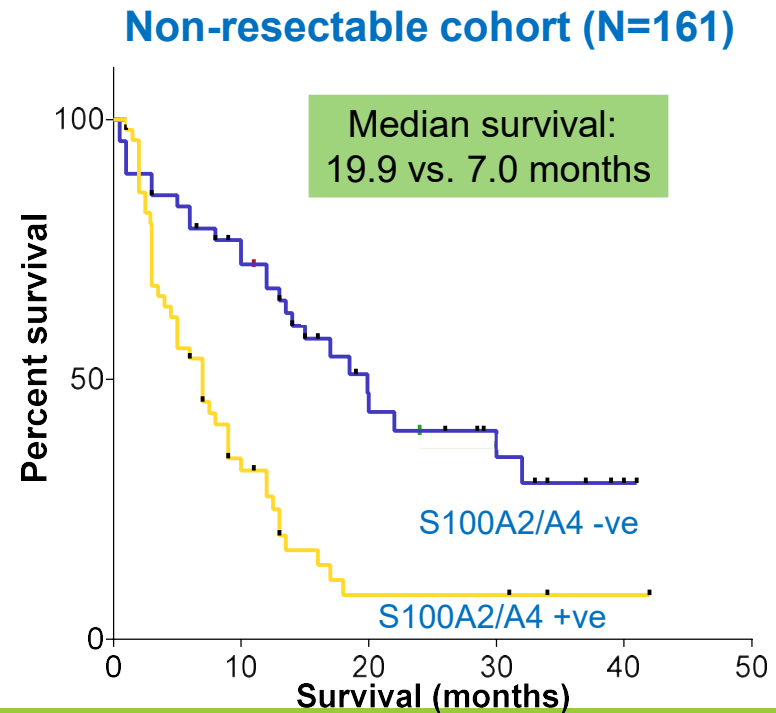
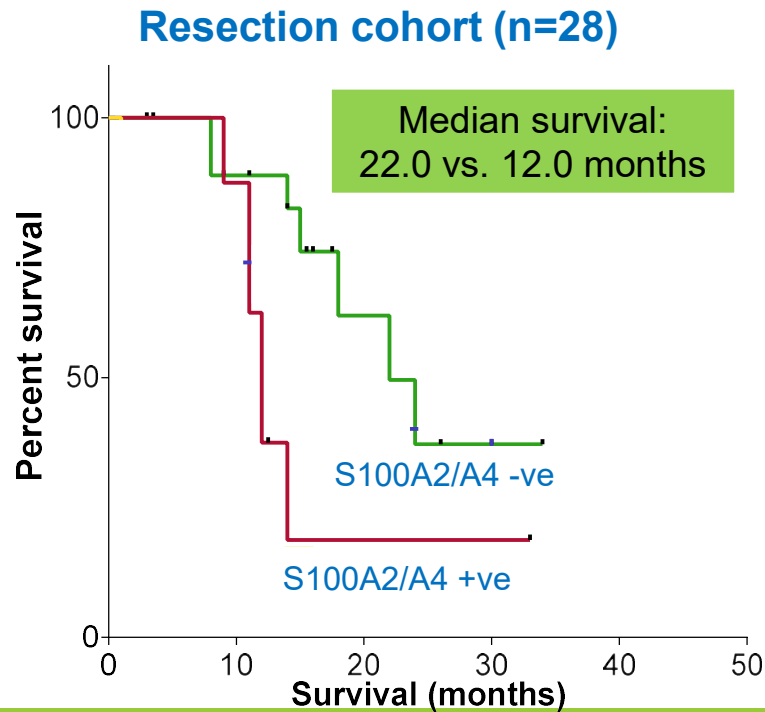
Inform outcomes and stratify appropriate therapy for patient



# IMPACT OF S100A2/4 ON OUTCOME OF PANCREATECTOMY

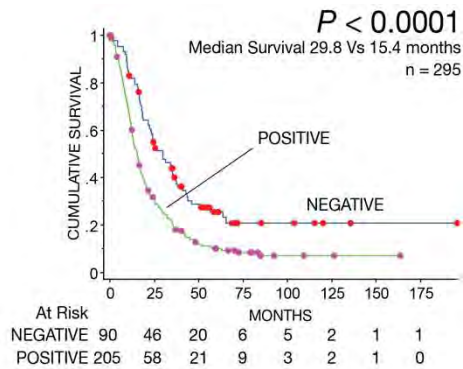
Prospective longitudinal study; 189 patients with pancreatic cancer  
EUS biopsy with FNB needle (22G, 20G PROCORE)

1. IHC studies (S100A2/A4)
2. Genomic analysis from micro-dissection of fresh tumor tissue

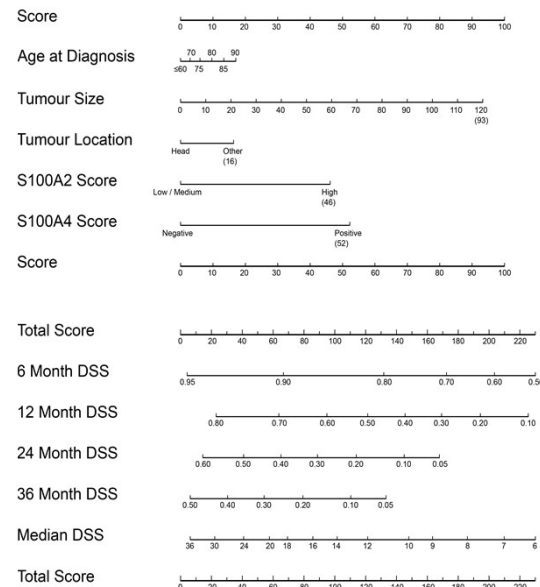


# Molecular nomogram to predict outcome of pancreatectomy

## S100A4

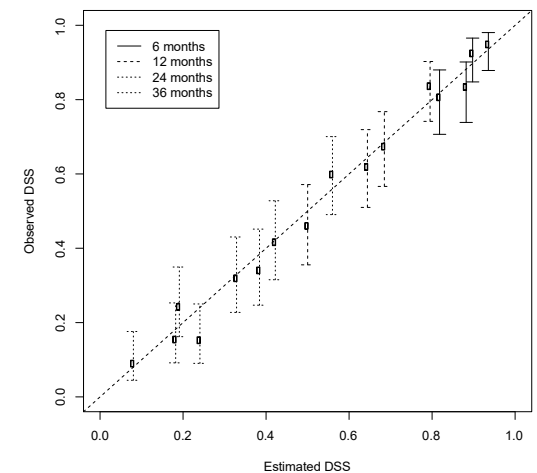
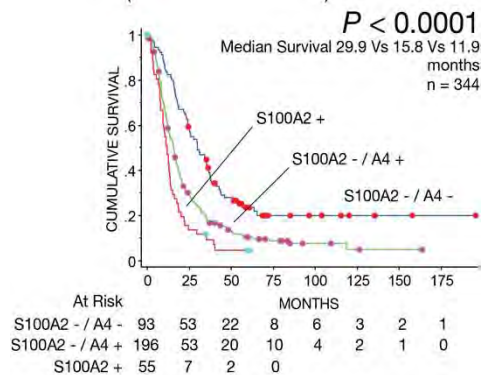


## Molecular Nomogram



Good correlation between the estimated and observed DSS

## S100A2 and A4

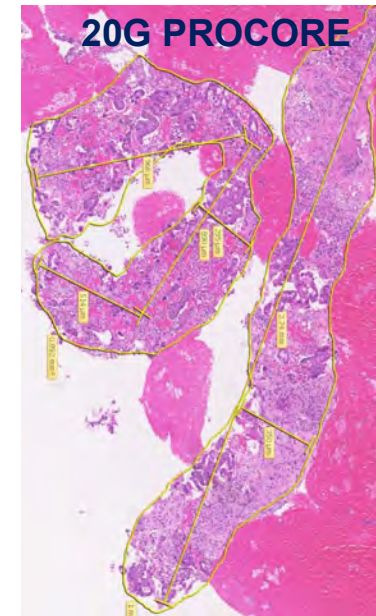
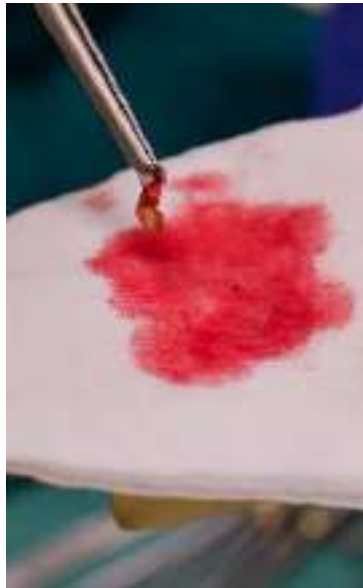


# How much tissue do I need to biomarker or genomic testing?

## Minimum requirement for genomic sequencing:

- 100 ng or more of total DNA
- ~500,000 *cancer* cells

- One microliter or 1 cubic mm of blood contains about 4 million red blood cells
- One microliter or 1 cubic mm of tissue contains about ~1-2 million cells



# Feasibility for Precision Oncology Care

| EUS needle type             | Size (G) | Fresh Frozen Specimen        |                              |
|-----------------------------|----------|------------------------------|------------------------------|
|                             |          | DNA yield [mean, range (ng)] | RNA yield [mean, range (ng)] |
| Boston Acquire <sup>®</sup> | 22       | 1,819 (133–7,350)            | 191 (30–1,187)               |
| Sharkcore <sup>®</sup>      | 19       | 2,170 (11.4–6,000)           | N/A                          |
| Sharkcore <sup>®</sup>      | 22       | 2,939 (1,134–7,595)          | 481 (40–1,790)               |
| Cook Procore <sup>®</sup>   | 20       | 1745 (290–4,750)             | 18 (3.6–44)                  |

| Patient cohort                 | Needle size (G) | Formalin fixed paraffin embedded specimen |                              |
|--------------------------------|-----------------|---|------------------------------|
|                                |                 | DNA yield [mean, range (ng)]              | RNA yield [mean, range (ng)] |
| Training set (n=14)            | 22              | 1,819 (133–7,350)                         | 191 (30–1,187)               |
| PRECISION-Panc EUS set (n=27)  | 22              | 2,694 (102–28,600)                        | N/A                          |
| PRECISION-Panc Core set (n=19) | Various         | 550 (0–1,730)                             |                              |

For next-generation sequence, a minimum of **100ng** of either DNA or RNA are required

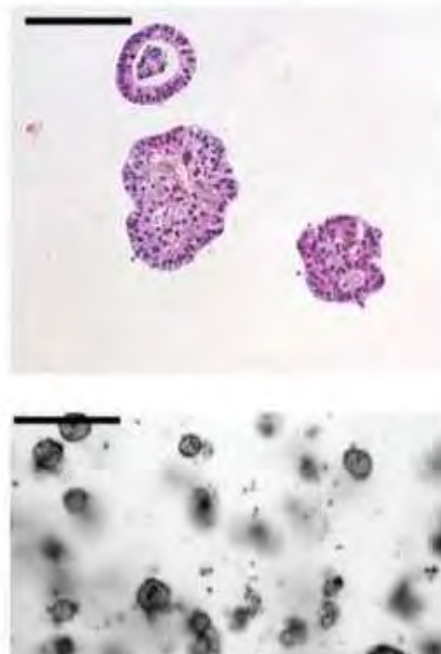
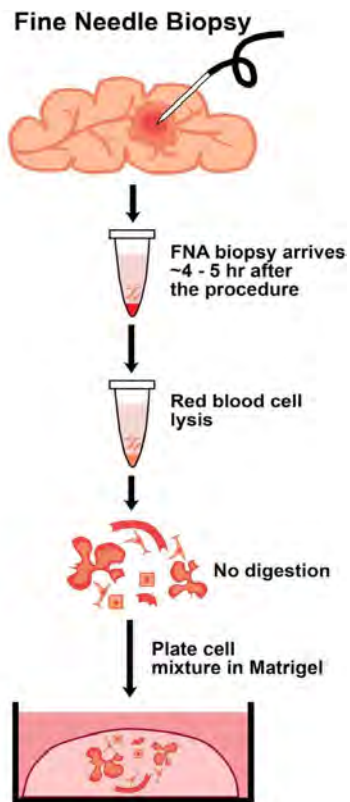
**31 of 43 patients (72%) had samples with sufficient quantity DNA for WGS**



# Genomic mutations to guide targeted therapy

| Target  | Treatment  |
|---|--|
| <i>KRAS</i> wild-type   | EGFR inhibitors (e.g., panitumumab, cetuximab, erlotinib)                          |
| DNA repair pathway defects ( <i>BRCA1</i> , <i>BRCA2</i> , <i>PALB2</i> , <i>ATM</i> )  | DNA damaging agents (e.g., mitomycin C, platinum) PARP inhibitors (e.g., olaparib) |
| <i>HER2</i> amplification   | Anti-HER2 antibodies/tyrosine kinase inhibitors (e.g., trastuzumab/lapatinib)      |
| <i>MET</i> activation (mutation, overexpression, amplification)                         | MET inhibitors   |
| Mismatch repair gene deficits ( <i>MLH1</i> , <i>MSH2</i> , <i>MSH6</i> , <i>PMS2</i> ) | Immunotherapy  |
| <i>PIK3CA</i> amplification/mutation +/- <i>PTEN</i> loss                               | mTOR inhibitors (e.g., everolimus)   |
| <i>CDKN2A</i> loss  | CDK4/6 inhibitors (e.g., palbociclib)  |
| <i>BRAF</i> mutation  | BRAF inhibitors (e.g., dabrafenib), MEK inhibitors (e.g., trametinib)              |
| <i>FGFR1</i> amplification  | FGFR inhibitors  |

# High feasibility of creating PDAC organoids from EUS FNB specimens



- N=38 histologically confirmed PDAC
- 2 extra passes of 22-gauge FNB needle (Procore)
- Within 2 weeks, isolation of **organoids** was achieved in 33 of 38 tumors (87%).
- Establishment of PDA **organoid** lines for  $\geq 5$  passages of growth (P5, five passages) was reached in 25 of 38 tumors (66%).
- There were no serious adverse events.

**Success rate is even better with the 20G Procore**  
(1 pass with 75% rate of  $>5$  passages, unpublished data)

# CONCLUSIONS

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The challenges to both endoscopist and pathologist for EUS guided biopsy can be overcome by advances in EUS-needle technology and appropriate post-FNA/B tissue processing

- *Data from RCTs indicate ROSE is not essential*
- *Direct histology processing has many advantages, especially for specimens taken from new core needles*
- *Routine use of FNB offer other advantages*
  - Increases specificity and improves inter-observer variability
  - Allow ancillary testing for Personalized Oncological treatment



**THANK YOU!**